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In Vivo Studies of various Antiparkinson’s agents: A Systematic Review

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Abstract: Parkinson’s disease (PD) is a neurodegenerative disorder which is characterized by typical symptoms including gradual progressive muscle rigidity, tremor and loss of motor skills. Although there is no definitive cure for PD, the extract of some medicinal plants and their ingredients have been suggested to relieve its symptoms and to prevent disability in patients. This review is focused on therapeutic effects of some anti-Parkinson’s agents. The findings presented in this review were collected from experimental studies in databases including PubMed, Web of Science and Google Scholar until the end of dec 2021. The keywords “Parkinson’s disease” or “neuroprotective” and “Medicinal plants”, “MPTP”, “6 OHDA”, “Rotenone”, and “haloperidol”, were searched. Based on the results of animal studies, according to animal model various anti Parkinson’s agents with their proposed mechanism are discussed in this review. This data will help to find new potential therapeutic agents for parkins disease treatment.

Keywords: Parkinson’s disease, Neuroprotective, MPTP, 6 OHDA, Rotenone, Haloperidol.

I. INTRODUCTION

Parkinson’s disease (PD) is primarily characterized by degradation of dopamine-carrying neurons in the substantia nigra with the extrapyramidal symptoms such as tremors, bradykinesia, rigidity, and inability to maintain the normal posture [1]. The neuronal death in PD is due to the damage to free radicals, Lewy’s bodies formation [2]. It has been prevalent in 10 million people around the globe with incidence rate of 219/100000 people in Pakistan [3]. There has been evidence that suggests the oxidative stress, accumulation of misfolded protein and the loss the dopaminergic neurons in substantia nigra pars compacta as the main hallmarks of PD pathogenesis [4]. The neurodegeneration has been accounted for the loss of 80% dopaminergic neurotransmission in striatum that leads to significant neuromuscular dysfunction along with some cognitive deficits at advanced stages [5]. Levodopa is the primary gold standard approach to symptomatically manage the PD but its chronic use has also been associated with development of dyskinesia [6]. Moreover, we have no therapeutic options that provides the neuroprotection or relieve the progression of PD. Therefore, it is the need of time to develop the therapeutic modalities that changes the course of PD progression along with treating it symptomatically.

One well-accepted and commonplace parkinsonian animal model is generated by intraperitoneal (i.p.) injection of 1-methyl-4-phenyl-1,2,3,6-tetrahydropyridine (MPTP), which is converted by monoamine oxidase type B (MAO-B) to its metabolite 1-methyl-4-phenylpyridinium (MPP+). MPP+ exhibits a high affinity for the dopamine transporter (DAT) and is transported into DA neurons, where it impairs respiration by inhibiting mitochondrial complex-1. This results in increased reactive oxygen species (ROS) production. ROS promotes cell death via oxidatively damaging molecules such as superoxide radicals and hydroxyl radicals and causes lipid and protein peroxidation. Eventually, the affected DA neurons can degenerate by either necrosis or apoptosis [7].

Table 1: Anti Parkinson’s agents effective in MPTP induced Parkinson’s disease.

Sr. No	Name	Parts and Family/ Extracts/Fraction	Maximum tolerated dose (MTD)/ Therapeutic doses (mg/kg)	Constituents/Possible responsible for this effect	mechanism
1.	(-) Epigallocatechin-3-gallate ^[8]		25 50	EGCG serves neuroprotective effects in an MPTP-induced PD mice model and may	

				exert this through modulating peripheral immune response.
2.	Acanthopanax senticosus harms ^[9]	Root and Rhizome Ethanol	182 45.5	It can increase the level of DA in striatum, balance the behavioral activation/inhibition at a striatal level and protect DA neurons from dying by apoptosis in Parkinson's disease mice.
3.	Apium graveolens L. ^[10]	Whole plants Chinese medicine methanolic	125, 250 375	Extract is able to ameliorate behavioral impairments, improve oxidative stress parameters, decrease the activity of MAO-A and B, and protect dopaminergic neurons.
4.	Apomorphine ^[11]		10	(a) its radical scavenging and iron-chelating properties; (b) its ability to protect against hydrogen peroxide, 6-OHDA, and iron-induced neurotoxicity in PC12 cell culture; (c) its ability to protect against MPTP induced neurotoxicity in vivo in mice; (d) its ability to inhibit mitochondrial iron-induced lipid peroxidation and protein oxidation; (e) its ability to prevent 6-OHDA induced inhibition of mitochondrial complex I activity; and (f) its ability to inhibit MAO-A and B.
5.	Chinonin ^[12]		10 20 40	Antioxidative property
6.	Cordycepin ^[13]		10 20	By inhibiting TLR/NF-κB signaling pathway
7.	Dendropanax morbiferus ^[14]	Leaves Aqueous	200	It effectively curbs the microglia-stimulated neuroinflammation by modulating the NF-κB/IκB-α and JNK-MAPK signaling pathways.
8.	Dexrazoxane ^[15]		1.5 5 15 3 10 30	Via attenuation of oxidative stress and ER stress, as well as the suppression of systemic inflammation in both peripheral tissues and brain.
9.	Entacapone ^[16]		12.5	To avoid pulsatile dopaminergic stimulation and provides increased therapeutic response without additional risk of dyskinesia induction over and above that found with a dopamine agonist alone.
10.	Eucommia ulmoides Oliv. ^[17]	Bark Ethanol	2.5 g 5 g 10 g	It is mediated by downregulating p38/JNK-Fosl2 gene expression to alleviate neuroinflammation.
11.	Eupatilin ^[18]		10	Inhibition of neuroinflammation and apoptosis associated with down-regulation

				of NF-κB signaling and up-regulation of Akt/GSK-3β signaling.
12.	Evernic Acid ^[19]		5 80	Neuroprotective and anti-inflammatory effects
13.	Fasudil ^[20]			Reduction of glial cell aggregation around the striatum and SN; Inhibition of ROCK expression; Induction of Nrf2/HO-1 antioxidant pathway; and regulation of NMDAR and AMPAR.
14.	Geniposide ^[21]		100	Geniposide exerted its neuroprotective effect by enhancing growth factor signaling and the reduction of apoptosis
15.	Gentisic acid ^[22]		80	Inhibition of oxidative stress
16.	Ginkgetin ^[23]		80 100	Neuroprotection and demonstrated its potential use as an antioxidant for the mitigation of PD.
17.	Isobavachalcone ^[24]		50 10	Isobavachalcone decreased the LPS-induced oxidative stress and the expression of inflammatory cytokines, and provided a neuroprotective effect by antagonizing microglia-mediated inflammation.
18.	Isolongifolene ^[25]		5, 10, 20	ILF showed the potential to arrest apoptosis through the inhibition of caspase activity and rebalancing of the Bax/Bcl-2 ratio in rotenone-treated rats.
19.	Lycopene ^[26]		5 10 20	Lycopene reverses neurochemical deficits, oxidative stress, apoptosis and physiological abnormalities in PD mice
20	Magnolol ^[27]		10 mL/kg	It may reverse the neuronal damage in the MPTP-lesioned PD mice.
21.	Norfluoxetine ^[28]		1 5 10	It is associated with neuroinflammation and microglia-derived oxidative stress.
22.	Ocimum sanctum ^[29]	Leaf Ethanollic	1.75, 4.25 8.5	Dopamine facilitatory and antioxidant properties.
23.	Oleanolic acid ^[30]			Inhibits the increase in reactive oxygen species which play a primary role in neurodegeneration in Parkinson's disease.
24.	Piperine ^[31]		10	Anti-apoptotic and anti-inflammatory mechanism on 6-OHDA induced Parkinson's disease
25.	Portulaca oleracea ^[32]	Seed Methanolic	2000/ 200 400	Inhibition of oxidative stress
26.	Quercetin ^[33]		30	Increased the activity of several antioxidant enzymes.

27.	Resveratrol ^[34]		10	By inhibiting neuro-inflammation, apoptosis and promoting neuronal survival.
28.	Rolipram ^[35]		3	By improving the cAMP/CREB signaling pathway in the DG.
29.	Silibinin ^[36]		280	On the one hand, silibinin alleviates mitochondrial damage in the brain of mice with Parkinson's disease by inhibiting oxidative stress, reducing inflammatory response and α -synuclein aggregation. On the other hand, silibinin can protect dopaminergic nerve by promoting mitophagy to remove damaged mitochondria in the brain of mice with Parkinson's disease
30.	Simvastatin ^[37]		1	Via inhibition of A1 reactive astrocytes in the MPTP mouse model of PD.
31.	Stemazole ^[38]		10 30 50	To enhanced anti-oxidative capacity, which may have resulted in the repair/restoration of dopaminergic neurons.
32.	Tripchlorolide ^[39]		1 μ g/kg	Marked increases in neurochemical and immunocytochemical indices of midbrain dopaminergic pathways by TW397 in MPTP parkinsonian model mice
33.	Troxaerutin ^[40]		150	Amelioration of apoptosis, astrogliosis, and oxidative stress and part of its effect is mediated through PI3K/ER β pathway.
34.	Ursolic acid ^[41]		5 10	The oxidative stress and inflammation triggered by rotenone was significantly diminished by UA.
35.	Uronic acid ^[42]		25	Inhibited MPP ⁺ -induced glial activation in primary astrocytes by blocking NF- κ B activation.
36.	Valerenic acid ^[43]		2	Inhibition of NF- κ B and activation of the 5-HT _{5A} receptor by its agonist valerenic acid in astrocytes.

6-Hydroxydopamine (6-OHDA) is a specific neurotoxin for catecholaminergic pathways (Perese et al. 1989; Sachs and Jonsson 1975). Being structurally similar to the catecholamines,

it uses the respective transport system to enter the neurons and destroys them. 6-OHDA has been reported to produce some of the behavioral, biochemical, and pathological changes that are encountered in Parkinson's disease (PD) (Bloem et al. 1990) and, because of established

stereotactic techniques and relatively low maintenance costs, is currently the most commonly used animal model for the disease (Breese and Breese 1998). These toxic effects of 6-OHDA are attributed to the formation of various oxidants and free radicals (Cohen 1984), lipid peroxidation (Slater 1984), protein damage, and amino acid modifications (Dean et al. 1985). In addition, studies have demonstrated that 6-OHDA leads to reduction in glutathione (GSH) content and superoxide dismutase (SOD) and catalase (CAT) activity, and an increase in lipid peroxidation (Perumal et al. 1992; Kumar et al. 1995; Zafar et al. 2003a, b; Ahmad et al. 2005a, b) in striatum.^[44]

Table 2: Anti Parkinson’s agents effective in 6-OHDA induced Parkinson’s disease.

Sr. No	Name	Parts and Family/ Extracts/ Fraction	Maximum tolerated dose (MTD)/ Therapeutic doses (mg/kg)	Constituents/Possible mechanism responsible for this effect
1.	(-)-sesamin ^[45]		30	Via the activation of transient ERK1/2- BadSer112 system and the inhibition of sustained ERK-p38MAPK-JNK1/2-caspase-3 system in PC12 cells. It showed prophylactic and adjuvant therapeutic effects on long-term L-DOPA therapy in dopaminergic neuronal cells of PD rat models.
2.	Albizia adianthifolia ^[46]	Leaves Aqueous	150 300	Antioxidant potential
3.	Bacopa monniera Linn ^[47]	Alcoholic	20 40	It has enhanced the availability of dopamine or might have prevented its breakdown and afford protection.
4.	Baicalein ^[48]	A flavonoid obtained from the root of Chinese medicinal herb Scutellaria baicalensis	200	By the increasing the number of dopaminergic neurons may have been, in part, caused by anti-apoptotic, pro differentiation and anti-inflammatory mechanisms of baicalein.
5.	Betaine ^[49]		12.5, 25, 50	Antioxidant and methyl donor properties of Betaine are promising particularly in management of plasma total homocysteine (tHcy) and oxidative stress in dopaminergic neurons of the brain.
6.	Caffeic Acid Phenethyl Ester ^[50]			The neuroprotective and anti-oxidant properties
7.	Caffeine and taurine ^[51]		10 8	It had an altering effect against the lesion induced by 6-OHDA as evaluated by behavioral tests and neurochemical analysis of striatal dopamine
8.	Cannabidiol ^[52]		10	The neuroprotective, anti-inflammatory and symptomatic effects of CBD treatment in an animal model of PD, potentially via the activation of astrocytic TRPV1-CNTF pathway.
9.	Cannabinoids ^[53]		3	These neuroprotective effects might be due, among others, to the antioxidant properties of certain plant-derived cannabinoids, or exerted through the capability of cannabinoid agonists to modulate glial function, or produced by a combination of both mechanisms.
10.	Cerebrolysin ^[54]		2.5ml/kg	Counteracting oxidative stress, replenishing dopamine content and enhancing behavioral outcomes.
11.	Curcumin and Desferrioxamine ^[55]		200 (curcumin) 50 (desferrioxamine)	Attenuated the loss of dopamine and increased antioxidant enzymes, resulting in preservation of dopaminergic neurons.
12.	Curcumin and naringenin ^[56]		50 (curcumin)	Antioxidant capabilities and their capability to penetrate into the brain.

			50(naringenin)	
13.	Curcumin ^[57]		5 10 20	Regulating intracellular bFGF/NGF/TrkA/Hsp70 expressions, thereby improving neurofunctions in the SN.
14.	Dexrazoxane ^[58]		1.5 5 15 3 10 30	Via attenuation of oxidative stress and ER stress, as well as the suppression of systemic inflammation in both peripheral tissues and brain.
15	Edaravone ^[59]		30 100 250	Anti-apoptotic effects and radical scavenging activity
16	Eugenol ^[60]		0.1, 1, 10	Improve the antioxidant response by increasing the production of GSH.
17	Fucoidan ^[61]		10 20	Suppress the Nox1-triggered oxidative stress in the SNc to protect DA neurons
18	Gallic acid ^[62]		50 100 200	GA has neuroprotective activity against 6-OHDA-induced oxidative stress <i>via</i> enhancement of cerebral antioxidant defense.
19	Garcinia indica ^[63]	Fruits Methanolic	100, 200, 400	Antioxidant and anti-inflammatory properties
20	Ginkgo biloba ^[64]		50 100 150	Ginkgo biloba appears to act <i>via</i> antioxidant, free radical scavenging, MAO-B-inhibiting, and DA-enhancing mechanisms that rescue the compromised cells within the dopaminergic lesions.
21	Gynostemma Pentaphyllum ^[65]	Leaves Ethanol	10 30	Protective effects against neurotoxicity by reducing TH neuronal cell death and normalizing dopamine levels in 6-OHDA-lesioned
22	Hemantane ^[66]		10	Possesses the antidyskinetic effect against the levodopa-induced dyskinesia disturbances
23	Hesperidin ^[67]		50	Increasing the DA levels, activity enzymatic and non enzymatic, decreasing the reactive species and improving the behavioral parameters
24	Hibiscus asper ^[68]	Leaves (Malvaceae) Methanolic	50 100	Antioxidant and antiapoptotic activities in Parkinson's disease model.
25	Humulus japonicas ^[69]	Cannabaceae	500	HJ improved the motor dysfunction and notably reduced dopaminergic cell death and fiber loss in the SNc and striatum caused by 6-OHDA.
26	Hypericum Perforatum ^[70]	Hydroalcoholic	200	Via attenuation of DNA fragmentation, astrogliosis, inflammation, and oxidative stress.
27	Montelukast ^[71]		10	A potential inhibitor of microglial activation to

			20 40	protect DA neurons in the adult brain against PD.
28	Mucuna pruriens [72]		2.5 5 g/kg	Mediated by increasing the mitochondrial complex-I activity and by scavenging the free radicals.
29	Naringenin [73]		70	To activate Nrf2/ARE pathway to add to the arsenal for treating neurodegenerative diseases
30	oleanolic acid [74]			Inhibits the increase in reactive oxygen species which play a primary role in neurodegeneration in Parkinson's disease.
31	Piperine [75]		10	Anti-apoptotic and anti-inflammatory mechanism on 6-OHDA induced Parkinson's disease
32	Quercetin [76]		30	Increased the activity of several antioxidant enzymes.
33	Sesame [77]	Seed Oil		By enhancing the activities of antioxidant enzymes, decreasing the TBARS content, TH positive expression and increased dopamine and its metabolite DOPAC level.
34	Sorafenib [78]		10	Maintain the normal range of natural antioxidant enzymes in brain tissue.
35	Stereospermum suaveolens DC [79]	Stem barks Methanolic	5000/ 125 250 500	It also contains saponin, a-cellulose, lignin, tannins, flavonoids, and saponins which may be responsible for the observed neuroprotective activity by direct antioxidant properties to detoxify ROS.
36	Syringic acid [80]		20	Via its neuroprotective, antioxidant and anti-inflammatory effects.
37	Thymol [81]		20 30 40	To an antioxidation mechanism
38	Thymoquinone [82]		5 10	Due to the attenuation of lipid peroxidation and this may provide benefits, along with other therapies,
39	Tinospora cordifolia [83]	Aerial parts Ethanol	200 400	By protecting dopaminergic neurons and reducing the iron accumulation.
40	Tricetin [84]			Protect dopaminergic neurons from 6-OHDA- induced neurotoxicity through mitochondrial apoptosis pathway and Nrf2/HO-1 signaling pathway.
41	Troxeutin [85]		150	Amelioration of apoptosis, astrogliosis, and oxidative stress and part of its effect is mediated through PI3K/ERβ pathway.
42	Vanillin [86]		20	Via preserving striatal dopamine levels
43	Varenicline [87]		1	Neuroprotective effect

Experimental as well as epidemiological studies provided evidence that exposure to many types of pesticides is accompanied by a greater risk of developing PD (Uversky et al. 2002; Uversky 2004). Rotenone is a pesticide and a potent inhibitor of complex I in the mitochondria (Naoi et al. 2005). Systemic administration of rotenone to rats produces nigrostriatal dopaminergic degeneration (Betarbet et al. 2000). Additionally, rotenone is well-characterized to be extremely hydrophobic and easily crosses biological membranes (Brown et al. 2006). Hence, rotenone model is a greatly reproducible tool for testing novel neuroprotective interventions for treating patients suffering from PD (Cannon et al. 2009).^[88]

Table 3: Anti Parkinson’s agents effective in Rotenone induced Parkinson’s disease.

Sr. No	Name	Parts and Family/ Extracts/Fraction	Maximum tolerated dose (MTD)/ Therapeutic doses (mg/kg)	Constituents/Possible mechanism responsible for this effect
1.	Agaricus Blazei Murill ^[89]	Aqueous	273 819	Decreasing oxidative stress in the animal brain by increasing the brain levels of reduced GSH and total proteins and decreasing the levels of nitrite and TBARS.
2.	Agomelatine ^[90]		40	Increased levels of caspase-3 expression propose apoptosis induced mechanism behind agomelatine induced neuronal loss.
3.	Boswellic acids ^[91]	Tablets	125, 250	To suppress pro-inflammatory cytokines and neurodegeneration
4.	Caffeic acid ^[92]		2.5, 5, 10	Anti-inflammatory activity of caffeic acid and highlighted its neuroprotective activity
5.	Carbenoxolone ^[93]		20	Prevents the mitochondrial dysfunctions and reduces the neuroinflammation caused by rotenone treatment.
6.	Crocin ^[94]		30	Via activation of PI3K/Akt/mTOR axis and enhanced miRNA-7 and miRNA-221.
7.	Demethoxycurcumin ^[95]		5, 10, 15	Its anti-inflammatory and antioxidant activities.
8.	Filgrastim ^[96]	Recombinant human G-CSF (filgrastim)	(20 and 40 µg/kg)	Reduction of rotenone-induced neuroinflammation, apoptosis, and brain-derived neurotrophic factor depletion
9.	Glycyrrhizic acid ^[97]		50	Its potent antioxidative and anti-inflammatory properties.
10	HidroX® ^[98]		10	Antioxidant, anti-inflammatory, prevented the α synuclein from aggregating and Forming accumulations in the

				dopaminergic neurons
11	Hyoscyamus niger ^[99]	Seeds Methanolic	125 250 500	Recovery in locomotor activity which may confer neuroprotection against the underlying dopaminergic neuron degeneration
12	Isolongifolene ^[100]		5, 10, 20	Anti-oxidant and antiapoptotic properties
13	Monascin ^[101]		100 200 400	Antioxidation and anti-neuroinflammation via modulating NF-κB and Nrf2 pathway.
14	Nerolidol ^[102]		50	Its antioxidant and anti-inflammatory activities.
15	Pomegranate Juice (Punica granatum L.) ^[103]			Its protection against oxidative damage and -synuclein aggregation, the increase in mitochondrial aldehyde dehydrogenase activity, and maintenance of antiapoptotic Bcl-xL protein at the control level.
16	Pulicaria undulate ^[104]	Essential oil	50 100 200	Anti-inflammatory and antioxidant activities with the ability to reduce a-synuclein gene expression
17	Sesaminol ^[105]			Reduces α-synuclein expression in the substantia nigra, which suppresses motor dysfunction and the decline of intestinal motor function.
18	Sida cordifolia ^[106]	Aqueous hexane (HFSC), chloroform (CFSC) and aqueous	50 100 250	Virtue of its antioxidative actions
19	Ursolic acid ^[107]		5 10	The oxidative stress and inflammation triggered by rotenone was significantly diminished by UA.
20	Vanillic acid ^[108]		12 25 50	Oxidative stress and attenuated the motor defects indicating the possible therapeutic potential of VA as a neuroprotective in PD.
21	Vitamin E ^[109]		100 I.U/Kg/ day i.m.	Potential antioxidant role of vitamin E in the nigrostriatal system.

Typical neuroleptic agents like chlorpromazine, haloperidol and reserpine induce a cataleptic state in rodents and these are being used as models to test the extrapyramidal side effects involved with it. Neuroleptic induced catalepsy has been linked to a blockade of postsynaptic striatal dopamine D1 and D2 receptors. Despite this evidence, several other neurotransmitters such as acetylcholine, serotonin, angiotensin, adenosine, or opioids have also been implicated. In addition to implications of various neurotransmitters in catalepsy, many preclinical and clinical studies have proposed reactive oxygen species in haloperidol induced toxicity. Evidence indicates that drugs which potentiate or attenuate neuroleptic catalepsy in rodents might aggravate or reduce the extrapyramidal signs respectively, in human beings.^[110]

Table 4: Anti Parkinson’s agents effective in Haloperidol induced Parkinson’s disease.

Sr. No	Name	Parts and Family/ Extracts/Fraction	Maximum tolerated dose (MTD)/ Therapeutic doses (mg/kg)	Constituents/Possible mechanism responsible for this effect
1.	<i>Achyranthes aspera</i> ^[111]	Whole plant Hydroalcoholic	2000/ 200 400	Possible antioxidant role of <i>A. aspera</i> extract in overcoming the neurochemical and behavioral changes during oxidative stress.
2.	<i>Albizia lebbek</i> (L.) ^[112]	Seeds Aqueous Methanolic	100 200 300	<i>Albizia lebbek</i> (L.) improved the motor functions and reversed the biochemical damages in brain tissue of PD
3.	<i>Beta vulgaris</i> L. ^[113]	Leaves (<i>Chenopodiaceae</i>) Methanolic	2000/ 100 200 300	Augmentation of cellular antioxidants
4.	<i>Brassica juncea</i> ^[114]	Leaves (<i>Cruciferae</i>)	200, 400, 600	<i>B. juncea</i> improved motor functions and enhanced the antioxidant enzymes in brain tissues. reduced the MAO-B levels in the brain
5.	Buspiron ^[115]		20	Activation of 5-HT1A receptors.
6.	<i>Cannabis sativa</i> ^[116]	Flowering tops and Leaves	5 10 20	<i>Cannabis</i> alters the oxidative status of the brain in favor of reducing lipid peroxidation, but reduces brain glucose, which would impair brain energetics.
7.	<i>Cucurbita pepo</i> ^[117]	Seeds (<i>Cucurbitaceae</i>) Methanolic	200 400 600	It has an antioxidant and neuroprotective effect due to phenols, flavonoids and beta-tocopherol
8.	<i>Cyamopsis tetragonoloba</i> ^[118]	Methanol	200 400	Antioxidant potential
9.	Dicyclomine ^[119]		40, 80, 160	Enhancement of antioxidant defense system
10.	<i>Elaeocarpus ganitrus</i> ^[120]	Elaeocarpaceae	100 200 400	It has anti-oxidant activity and neuroprotective activity
11	<i>Emblica officinalis</i> ^[121]	Fruit Aqueous	0.8, 2.0 4.0	Due to both its anticholinergic and antioxidant properties.
12	<i>Euphorbia cyathophora</i> ^[122]	Leaves Ethanolic	2000/ 200, 400	Attenuated the motor defects and also increased the neuro chemical dopamine level.
13	<i>Ficus religiosa</i> ^[123]	Leaves Petroleum ether	4000/ 100, 200, 400	<i>Ficus religiosa</i> treatment significantly attenuated the motor defects and also protected the brain from oxidative stress.
14	Flupirtine ^[124]		1 10	It synergises with dopaminomimetics, it may prevent development of L-DOPA-induced fluctuations as

			20	NMDA receptor antagonists do _Chase et al., 1996., it possesses a neuroprotective potential, and _iv. it is devoid of the side effects of NMDA receptor antagonists.
15	Gentisic acid ^[125]		80	Inhibition of oxidative stress
16	Glucocorticoids ^[126]		1 2	Anticataleptic action of glucocorticoids.
17	Green coffee extract ^[127]	Seeds	100 400	Indirectly modulate dopaminergic transmission
18	Lauric acid ^[128]		0.66 1.32	Neuro-protection against oxidative stress, inflammatory cytokines and behavioral changes in HPD induced rat model of PkD.
19	Metformin ^[129]		25 50 100	Inhibition of oxidative/nitrosative stress
20	Myrica esulenta ^[130]	Leaves Methanolic	2000/ 50 100 200	Due to an escalation of cellular antioxidants
21	Nardostachys jatamansi ^[131]	Dried roots Aqueous	5000 250 500	Antioxidant potential has contributed to the reduction in the oxidative stress and catalepsy induced by haloperidol administration.
22	Ocimum sanctum ^[132]	Leaf Ethanollic	1.75, 4.25 8.5	Dopamine facilitatory and antioxidant properties.
23	Phaseolus vulgaris ^[133]	Seeds Methanolic	200	Herb contains L-DOPA and also possess the antioxidant activity.
24	Portulaca oleracea ^[134]	Seed Methanolic	2000/ 200 400	Inhibition of oxidative stress
25	Rhinacanthin-C ^[135]		5 10 20	The compound improves catalepsy and locomotion by increasing dopamine, serotonin, and norepinephrine concentration in the brain.
26	Tribulus terrestris ^[136]	Fruits Methanol	100 300 1000	Modulation of AChE, α -Synuclein, TNF- α , and IL-1 β
27	Tridax procumbens ^[137]	Leaves (Asteraceae) Ethanollic	100 200	Due to its neuroprotective and free radical scavenging properties.
28	Varenicline ^[138]		0.5 1.5 2.5	It can delay the rate of progression of PD, but also alleviates the symptoms of PD.
29	Vigna aconitifolia ^[139]	Seeds Hydroalcoholic	2000/ 100 200 300	The predictable mode of action of this plant may be due to increased synthesis of dopamine from L-dopa and decreased lipid peroxidation due to the presence of flavonoids and polyphenols.
30	Withania somnifera ^[140]	Root	1.7, 4.25, 8.5	Antioxidant properties

Table 4.1: Anti Parkinson’s agents effective in chlorpromazine induced Parkinson’s disease.

Sr. No	Name	Parts and Family/ Extracts/Fraction	Maximum tolerated dose (MTD)/ Therapeutic doses (mg/kg)	Constituents/Possible mechanism responsible for this effect
1	Camel milk ^[141]		33ml/kg p.o	Neuroprotective effect of camel milk could be attributed to its antioxidant property.
2	Diclofenac ^[142]		20	Via preventing dopaminergic neuronal cell death
3	Phaseolus vulgaris ^[143]	Seeds Methanolic	200	Presence of L-dopa in <i>Phaseolus vulgaris</i> in phytochemical screening of herb

REFERENCES

[1] Vegeto E, Benedusi V and Maggi A. Estrogen Anti-Inflammatory Activity in Brain: A Therapeutic Opportunity for Menopause and Neurodegenerative Diseases. *Front Neuroendocrinol* 2008; 29(4):507-19.

[2] Shibeshi W, Makonnen E, Zerihun L and Debella A: Effect Of *Achyranthes Aspera* L. On Fetal Abortion, Uterine and Pituitary Weights, Serum Lipids and Hormones. *Afr Health Sci* 2006; 6(2):108-12.

[3] Hussain G, Rasul A, Anwar H, Sohail Mu, Kashif Shahid Kamran S, Baig S, Shabbir A, Iqbal J: Epidemiological Data Of Neurological Disorders In Pakistan And Neighboring Countries: A Review. *Pakistan. J Neurol Sci* 2017; 12(4):12.

[4] Kaur R, Mehan S, Singh S: Understanding Multifactorial Architecture of Parkinson's Disease: Pathophysiology To Management. *Neurol Sci* 2019; 40(1):13–23.

[5] Cheng H-C, Ulane Cm, Burke Re: Clinical Progression in Parkinson Disease and the Neurobiology of Axons. *Ann Neurol* 2010; 67(6):715–25.

[6] Thanvi B, Lo N, Robinson T: Levodopa-Induced Dyskinesia In Parkinson's Disease: Clinical Features, Pathogenesis, Prevention And Treatment. *Postgrad Med J* 2007; 83(980):384–8.

[7] Pennapa Chonpathompikunlert, Phetcharat Boonruamkaew, Wanida Sukketsiri, Pilaiwanwadee Hutamekalin and Morakot Sroyraya: The Antioxidant and Neurochemical Activity of *Apium Graveolens* L. And Its Ameliorative Effect on Mptp-Induced Parkinson-Like Symptoms in Mice. *Bmc Complementary and Alternative Medicine* 2018; 103(18): 1-12.

[8] Tingting Zhou, Mengru Zhu and Zhanhua Liang: (-)-Epigallocatechin-3-Gallate Modulates Peripheral Immunity in The Mptp-Induced Mouse Model of Parkinson's Disease. *Molecular Medicine Reports* 2018; 17: 4883-4888.

[9] Shu-Min Liu, Xu-Zhao Li, Yan Huo, Fang Lu: Protective Effect Of Extract Of *Acanthopanax Senticosus* Harms On Dopaminergic Neurons In Parkinson's Disease Mice. *Phytomedicine* 2012; 19:631–638.

[10] Pennapa Chonpathompikunlert, Phetcharat Boonruamkaew, Wanida Sukketsiri, Pilaiwanwadee Hutamekalin and Morakot Sroyraya: The Antioxidant and Neurochemical Activity of *Apium Graveolens* L. And Its Ameliorative Effect on Mptp-Induced Parkinson-Like Symptoms in Mice. *Bmc Complementary and Alternative Medicine* 2018; 18(103):1-12.

[11] Edna Grunblatt, Dsc (Student), Silvia Mandel, Dsc, Tamara Berkuzki, Dsc, And M. B. H. Youdim, Prof Phd: Apomorphine Protects Against Mptp-Induced Neurotoxicity in Mice. *Movement Disorders* 1999; 14(4): 612–618.

[12] Guoshuai Feng, Zhijian Zhang, Qingqing Bao, Zaijun Zhang, Libing Zhou, Jie Jiang, And Sha Li: Protective Effect Of Chinonin In Mptp-Induced C57bl/6 Mouse Model Of Parkinson's Disease. *Biol. Pharm. Bull* 2014; 37(8): 1301–1307.

[13] Chunyan Cheng, Xiaoying Zhu: Cordycepin Mitigates Mptp-Induced Parkinson's Disease Through Inhibiting Tlr/Nf-Kb Signaling Pathway. *Life Sci* 2019; 223:120-127.

[14] Shin-Young Park, Govindarajan Karthivashan , Hyun Myung Ko , Duk-Yeon Cho, Joonsoo Kim, Dae Jun Cho, Palanivel Ganesan, In Su-Kim, And Dong-Kug Choi: Aqueous Extract Of *Dendropanax Morbiferus* Leaves Effectively Alleviated Neuroinflammation And Behavioral Impediments In Mptp-Induced Parkinson's Mouse Model. *Oxidative Medicine and Cellular Longevity* 2018; 3175214: 1-14.

[15] Meng Mei , Yuanzhang Zhou , Mengdi Liu , Fangfang Zhao , Cong Wang , Jianhua Ding , Ming Lu, Gang Hu: Antioxidant And Anti-Inflammatory Effects Of Dextrazoxane On Dopaminergic Neuron Degeneration In Rodent Models Of Parkinson's Disease. *Neuropharmacology* 2019; 107758(160): 1-11.

[16] Mohammed Zubair, Michael J. Jackson, Kayhan Tayarani-Binazir, Kim A. Stockwell, Lance A. Smith, Sarah Rose, Warren Olanow, and Peter Jenner: The Administration Of Entacapone Prevents L-Dopa-Induced Dyskinesia When Added To Dopamine Agonist Therapy In Mptp-Treated Primates. *Experimental Neurology* 2007; 208:177–184.

[17] Shanshan Fan, Qingsheng Yin, Dongna Li, Jing Ma, Lili Li, Shiwei Chai, Hong Guo, Zhen Yang: Anti-Neuroinflammatory Effects Of *Eucommia Ulmoides* Oliv. In A Parkinson's Mouse Model Through the Regulation of P38/Jnk-Fosl2 Gene Expression. *Journal of Ethnopharmacology* 2020; 260: 1-43.

- [18] Yuan Zhang, Lina Qin, Jieru Xie, Juan Li, Chong Wang: Eupatilin Prevents Behavioral Deficits and Dopaminergic Neuron Degeneration in A Parkinson's Disease Mouse Model. *Life Sciences* 2020; 253(117745): 1-9.
- [19] Seulah Lee, Yeon Ji Suh, Seonguk Yang, Dong Geun Hong, Akihito Ishigami, Hangun Kim, Jae-Seoun Hur, Seung-Cheol Chang And Jaewon Lee: Neuroprotective And Anti-Inflammatory Effects Of Evermic Acid In An Mptp-Induced Parkinson's Disease Model. *Int. J. Mol. Sci* 2021; 22(2098): 1-18.
- [20] Yan-Hua Li, Jing-Wen Yu, Jian-Yin Xi, Wen-Bo Yu, Jian-Chun Liu, Qing Wang, Li-Juan Song, Ling Feng, Ya-Ping Yan, Guang-Xian Zhang, Bao-Guo Xiao, Cun-Gen Ma: Fasudil Enhances Therapeutic Efficacy Of Neural Stem Cells In The Mouse Model Of Mptp-Induced Parkinson's Disease. *Mol Neurobiol* 2016: 1-14.
- [21] Yimei Chen, Yanfang Zhang, Lin Li, Christian Hölscher: Neuroprotective Effects of Geniposide in The Mptp Mouse Model of Parkinson's Disease. *Eur J Pharmacol* 2015; 768:21-7.
- [22] Kabra M , Bhandari Ss, Sharma A, Gupta Rb: Evaluation Of Anti-Parkinson's Activity Of Gentisic Acid In Different Animal Models. *Journal of Acute Disease* 2014:141-144.
- [23] Y-Q Wang, M-Y Wang, X-R Fu, Peng-Yu, G-F Gao, Y-M Fan, X-L Duan, B-L Zhao, Y-Z Chang, Z-H Shi: Neuroprotective Effects Of Ginkgetin Against Neuroinjury In Parkinson's Disease Model Induced By Mptp Via Chelating Iron. *Free Radic Res* 2015; 49(9):1069-80.
- [24] Haoran Jing, Shaoxia Wang, Min Wang, Wenliang Fu, Chao Zhang, Donggang Xu: Isobavachalcone Attenuates Mptp-Induced Parkinson's Disease In Mice By Inhibition Of Microglial Activation Through Nf-Kb Pathway. *Plos One* 2017: 1-15.
- [25] Rengasamy Balakrishnan, Dhanraj Vijayaraja, Thangavel Mohankumar, Dharmar Manimaran, Palanivel Ganesan, Dong-Kug Choi, Namasivayam Elangovan: Isolongifolene Mitigates Rotenone-Induced Dopamine Depletion And Motor Deficits Through Anti-Oxidative And Anti-Apoptotic Effects In A Rat Model Of Parkinson's Disease. *Journal of Chemical Neuroanatomy* 2021; 112(101890): 1-12.
- [26] Asokan Prema, Udaiyappan Janakiraman, Thamilarasan Manivasagam, Arokiasamy Justin Thenmozhi: Neuroprotective Effect of Lycopene against Mptp Induced Experimental Parkinson's Disease in Mice. *Neurosci Lett* 2015; 599: 12-9.
- [27] Chi-Chang Weng, Zi-An Chen, Ko-Ting Chao, Ting-Wei Ee, Kun-Ju Lin, Minghuan Chan, Ing-Tsung Hsiao, Tzu-Chen Yen, Mei-Ping Kung, Ching-Han Hsu, Shiau-Pyng Wey: Quantitative Analysis Of The Therapeutic Effect Of Magnolol On Mptp-Induced Mouse Model Of Parkinson's Disease Using In Vivo 18f-9-Fluoropropyl-(+)-Dihydrotetrabenazine Pet Imaging. *Plos One* 2017; 12(3):1-13.
- [28] Kyung In Kim, Young Cheul Chung , And Byung Kwan Jin: Norfluoxetine Prevents Degeneration Of Dopamine Neurons By Inhibiting Microglia-Derived Oxidative Stress In An Mptp Mouse Model Of Parkinson's Disease. *Mediators of Inflammation* 2018; 4591289: 1-8.
- [29] S. Pemminati, V. Nair, P. Dorababu, H.N. Gopalakrishna, M.R.S.M. Pai: Effect of Ethanolic Leaf Extract of Ocimum Sanctum on Haloperidol-Induced Catalepsy in Albino Mice. *Indian J Pharmacol* 2007; 39(2): 87-89.
- [30] Musa V. Mabandla, Mpumelelo Nyoka, Willie M.U. Daniels: Early Use Of Oleanolic Acid Provides Protection Against 6-Hydroxydopamine Induced Dopamine Neurodegeneration. *Brain Research* 2015; 1622:64-71.
- [31] Pallavi Shrivastava, Kumar Vaibhav, Rizwana Tabassum, Andleeb Khan, Tauheed Ishrat, Mohd. Moshahid Khan, Ajmal Ahmad, Farah Islam, Mohammed M. Safhi, Fakhru Islam: Anti-Apoptotic And Anti-Inflammatory Effect Of Piperine On 6-OHda Induced Parkinson's Rat Model. *J Nutr Biochem* 2013; 24(4):680-7.
- [32] Santosh Kumar Vaidya, Dharmesh K. Golwala, Darpini S. Patel and Satyajit Sahoo: Evaluation of Antioxidant and Anti-Parkinson Activity of *Portulaca Oleracea* Seed Methanolic Extract. *Ejmp* 2020; 31(2): 10-17.
- [33] Nagaraja Haleagrahara, Cheng Jun Siew, Nilesh Kumar Mitra, Mangala Kumari: Neuroprotective Effect of Bioflavonoid Quercetin in 6-Hydroxydopamine-Induced Oxidative Stress Biomarkers in the Rat Striatum. *Neuroscience Letters* 2011; 500:139-143.
- [34] Qianqian Liu, Dashuai Zhu, Peien Jiang, Xinyu Tang , Qiuhuan Lang , Qinyi Yu , Shaozhi Zhang, Yongzhe Che , Xizeng Feng: Resveratrol Synergizes With Low Doses Of L-Dopa To Improve Mptp-Induced Parkinson Disease In Mice. *Behavioural Brain Research* 2019; 367:10-18.
- [35] Ken-Ichi Kinoshita, Yoshikage Muroi, Toshihiro Unno, Toshiaki Ishii: Rolipram Improves Facilitation Of Contextual Fear Extinction In The 1-Methyl-4-Phenyl-1,2,3,6-Tetrahydropyridine-Induced Mouse Model Of Parkinson's Disease. *J Pharmacol Sci* 2017; 134(1):55-58.
- [36] Xiumin Liu , Weiwei Liu , Chenkang Wang , Yinzhe Chen , Panwen Liu , Toshihiko Hayashi, Kazunori Mizuno , Shunji Hattori , Hitomi Fujisaki , Takashi Ikejima: Silibinin Attenuates Motor Dysfunction In A Mouse Model Of Parkinson's Disease By Suppression Of Oxidative Stress And Neuroinflammation Along With Promotion Of Mitophagy. *Physiology & Behavior* 2021; 239(113510):1-13.
- [37] Ren-Wei Du Wen-Guang Bu: Simvastatin Prevents Neurodegeneration in the Mptp Mouse Model of Parkinson's Disease via Inhibition of A1 Reactive Astrocytes. *Neuroimmunomodulation* 2021; 28:82-89.
- [38] Zhirui Guo, Shasha Xu, Na Du, Jia Liu, Yiyun Huang , Mei Han: Neuroprotective Effects Of Stemazole In The Mptp-Induced Acute Model Of Parkinson's Disease: Involvement Of The Dopamine System. *Neuroscience Letters* 2016; 616:152-159.
- [39] Zhen Hong, Gang Wang, Jing Gu, Jing Pan, Li Bai and Shi Zhang, Sheng-Di Chen: Triphlorolide Protects Against Mptp-Induced Neurotoxicity in C57bl/6 Mice. *European Journal of Neuroscience* 2007; 26:1500-1508.
- [40] Tourandokht Baluchnejadmojarad, Nida Jamali-Raeufy, Sedigheh Zabihnejad , Nafiseh Rabiee, Mehrdad Roghani: Troxerutin Exerts Neuroprotection In 6-Hydroxydopamine Lesion Rat Model Of Parkinson's Disease: Possible Involvement Of Pi3k/Erβ Signaling. *Eur J Pharmacol* 2017; 801:72-78.
- [41] Vaibhavi Peshattiwari, Suraj Muke, Aakruti Kaikini, Sneha Bagle, Vikas Dighe, Sadhana Sathaye: Mechanistic Evaluation Of Ursolic Acid Against Rotenone Induced Parkinson's Disease- Emphasizing The Role Of Mitochondrial Biogenesis. *Brain Res Bull* 2020; 160:150-161.
- [42] Seulah Lee, Yujeong Lee, Sugyeong Ha, Hae Young Chung, Hangun Kim, Jaeseoun Hur, Jaewon Lee: Anti-Inflammatory Effects Of Usnic Acid In An Mptp-Induced Mouse Model Of Parkinson's Disease. *Brain Res* 2020; 1730(146642): 1-38.
- [43] Alfredo Rodríguez-Cruz , Antonio Romo-Mancillas, Jesus Mendiola-Precoma, Jesica Esther Escobar-Cabrera, Guadalupe García-Alcocer, Laura Cristina Berumen: Effect Of Valerenic Acid On Neuroinflammation In A Mptp-Induced Mouse Model Of Parkinson's Disease. *Ibro Reports* 2020; 8:28-35.
- [44] Chandrasekar Shobana, Radhakrishnan Ramesh Kumar, Thangarajan Sumathi: Alcoholic Extract of *Bacopa Monniera* Linn. Protects Against 6-Hydroxydopamine-Induced Changes in Behavioral and Biochemical Aspects: A Pilot Study. *Cell Mol Neurobiol* 2012; 32:1099-1112.
- [45] Hyun Jin Park, Ting Ting Zhao, Kyung Sook Lee, Seung Ho Lee, Keon Sung Shin, Keun Hong Park, Hyun Sook Choi, Myung Koo Lee: Effects of (-)-Sesamin On 6-Hydroxydopamine-Induced Neurotoxicity In Pc12 Cells And Dopaminergic Neuronal Cells Of Parkinson's Disease Rat Models. *Neurochemistry International* 2015; 83-84.

- [46] Galba Jean Beppe, Alain Bertrand Dongmo, Harquin Simplicie Foyet, Nolé Tsabang, Zenovia Olteanu, Oana Cioanca, Monica Hancianu, Théophile Dimo And Lucian Hritcu: Memory-Enhancing Activities Of The Aqueous Extract Of Albizia Adianthifolia Leaves In The 6-Hydroxydopamine-Lesion Rodent Model Of Parkinson's Disease. *Bmc Complementary and Alternative Medicine* 2014; 142(14):1-11.
- [47] Chandrasekar Shobana, Radhakrishnan Ramesh Kumar, Thangarajan Sumathi: Alcoholic Extract of Bacopa Monniera Linn. Protects Against 6-Hydroxydopamine-Induced Changes in Behavioral and Biochemical Aspects: A Pilot Study. *Cell Mol Neurobiol* 2012; 32:1099-1112.
- [48] Xin Mu, Guorong He, Yinxia Cheng, Xiaoxiu Li, Bei Xu, Guanhua Du: Baicalein Exerts Neuroprotective Effects In 6-Hydroxydopamine-Induced Experimental Parkinsonism In Vivo And In Vitro. *Pharmacology, Biochemistry and Behavior* 2009; 92:642-648.
- [49] Milind V. Masule, Sachin D. Shinde, Sujata S. Kurkute, Balu U. Salve: Evaluation Of Antioxidant And Anti Parkinsonism Activity Of Betaine In Experimental Rats. *Journal of Drug Delivery & Therapeutics* 2019; 9(2-S):427-421.
- [50] Duygu Turan, Hüseyin Abdik, Fikrettin Şahin, Ezgi Avcı: Evaluation Of The Neuroprotective Potential Of Caffeic Acid Phenethyl Ester In A Cellular Model Of Parkinson's Disease. *Eur J Pharmacol* 2020; 883:1-38.
- [51] Amjad N. Abuirmeleh, Sawsan M. Abuhamdah, Asser Ashrafa And Kareem H. Alzoubi: Protective Effect Of Caffeine And/Or Taurine On The 6-Hydroxydopamine-Induced Rat Model Of Parkinson's Disease: Behavioral And Neurochemical Evidence. *Restorative Neurology and Neuroscience* 2021; 39:149-157.
- [52] Claudio Giuliano, Miriam Francavilla, Gerardo Ongari, Alessandro Petese, Cristina Ghezzi, Nora Rossini, Fabio Blandini, And Silvia Cerri: Neuroprotective And Symptomatic Effects Of Cannabidiol In An Animal Model Of Parkinson's Disease. *Int. J. Mol. Sci* 2021; 8920(22):1-14.
- [53] Isabel Lastres-Becker, Francisco Molina-Holgado, Jose´ A. Ramos, Raphael Mechoulam, And Javier Ferna Ndez-Ruiz: Cannabinoids Provide Neuroprotection Against 6-Hydroxydopamine Toxicity In Vivo And In Vitro: Relevance To Parkinson's Disease. *Neurobiology of Disease* 2005; 19:96-107.
- [54] Neveen A. Noor, Haitham S. Mohammed, Iman M. Mourad, Yasser A. Khadrawy, Heba S. Aboul Ez: A Promising Therapeutic Potential Of Cerebroslyn In 6-Ohda Rat Model Of Parkinson's Diseas. *Life Sci* 2016; 155:174-9.
- [55] Hua Lv, Jun Liu, Li Wang, Hong Zhang, Shuqi Yu, Zhiwei Li, Feng Jiang, Yu Niu, Jie Yuan, Xiaoli Cui, Wenxiu Wang: Ameliorating Effects Of Combined Curcumin And Desferrioxamine On 6-Ohda-Induced Rat Mode Of Parkinson's Disease. *Cell Biochem Biophys* 2014; 70:1433-1438.
- [56] Virginia Zbarsky, Krishna P. Datla, Shabnam Parkar, Deepal K. Rai, Okezie I. Aruoma, & David T. Dexter: Neuroprotective Properties Of The Natural Phenolic Antioxidants Curcumin And Naringenin But Not Quercetin And Fisetin In A 6-Ohda Model Of Parkinson's Disease. *Free Radical Research* 2005; 39(10):1119-1125.
- [57] Shilei Song Qingmei Nie Zhifang Li Gang Du: Curcumin Improves Neurofunctions Of 6-Ohda-Induced Parkinsonian Rats. *Pathol Res Pract* 2016; 212(4):247-51.
- [58] Meng Mei, Yuanzhang Zhou, Mengdi Liu, Fangfang Zhao, Cong Wang, Jianhua Ding, Ming Lu, Gang Hu: Antioxidant And Anti-Inflammatory Effects Of Dexrazoxane On Dopaminergic Neuron Degeneration In Rodent Models Of Parkinson's Disease. *Neuropharmacology* 2019; 160(107758):1-11.
- [59] Wen Ji Yuan, Takao Yasuhara, Tetsuro Shingo, Kenichiro Muraoka, Takashi Agari, Masahiro Kameda, Takashi Uozumi, Naoki Tajiri, Takamasa Morimoto, Meng Jing, Tanefumi Baba, Feifei Wang, Hanbai Leung, Toshihiro Matsui, Yasuyuki Miyoshi and Isao Date: Neuroprotective Effects Of Edaravone-Administration On 6-Ohda-Treated Dopaminergic Neurons. *Bmc Neuroscience* 2008; 9(75):1-11.
- [60] Carlos Franciney Moreira Vasconcelos, Nívea Maria Da Cunha Ferreira, Nayan Hardy Lima Pontes, Thomas Dominik De Sousa Dos Reis, Ricardo Basto Souza, Francisco Eduardo Aragão Catunda Junior, Lissiana Magna Vasconcelos Aguiar, Rodrigo Maranguape Silva Da Cunha: Eugenol And Its Association With Levodopa In 6-Hydroxydopamine-Induced Hemiparkinsonian Rats: Behavioural And Neurochemical Alterations. *Basic Clin Pharmacol Toxicol* 2020; 127(4):287-302.
- [61] Fei-Long Zhang, Yi He, Yan Zheng, Wen-Jing Zhang, Qi Wang, Yan-Jun Jia, Hai-Long Song, Hai-Ting An, Hao-Bo Zhang, Yan-Jing Qian, Yu-Long Tong, Lin Dong & Xiao-Min Wang: Therapeutic Effects Of Fucoidan In 6-Hydroxydopamine-Lesioned Rat Model Of Parkinson's Disease: Role Of Nadph Oxidase-1. *Cns Neuroscience & Therapeutics* 2014; 20:1036-1044.
- [62] Mohammad Taghi Mansouri, Yaghoob Farbood, Maryam Jafar Sameri, Alireza Sarkaki, Bahareh Naghizadeh, Maryam Rafeirad: Neuroprotective Effects Of Oral Gallic Acid Against Oxidative Stress Induced By 6- Hydroxydopamine In Rats. *Food Chem* 2013; 138(2-3):1028-33.
- [63] Bhaveshkumar V. Antala, Manishkumar S. Patel, Satish V. Bhuva1, Shiv Gupta2, Samir Rabadiya3, Mangala Lahkar: Protective Effect Of Methanolic Extract Of Garcinia Indica Fruits In 6-Ohda Rat Model Of Parkinson's Disease. *Indian Journal of Pharmacology* 2012; 44(6):683-687.
- [64] Muzamil Ahmad, Sofiyah Saleem, Abdullah Shafique Ahmad, Seema Yousuf, Mubeen Ahmad Ansari, M Badruzzaman Khan, Tauheed Ishrat, Rajnish Kumar Chaturvedi, Ashok Kumar Agrawal And Fakhru Islam: Ginkgo Biloba Affords Dose-Dependent Protection Against 6-Hydroxydopamine-Induced Parkinsonism In Rats: Neurobehavioural, Neurochemical And Immunohistochemical Evidences. *Journal of Neurochemistry* 2005; 93:94-104.
- [65] Hyun Sook Choi, Mi Sook Park, Seung Hwan Kim, Bang Yeon Hwang, Chong Kil Lee and Myung Koo Lee: Neuroprotective Effects of Herbal Ethanol Extracts from Gynostemma Pentaphyllum in the 6-Hydroxydopamine-Lesioned Rat Model of Parkinson's Disease. *Molecules* 2010; 15:2814-2824.
- [66] E. A. Ivanova, I. G. Kapitsa, E. A. Val'dman, And T. A. Voronina: Anti-Parkinsonian Activity Of Hemantane On A Model Of Hemiparkinsonian Syndrome In Rats. *Bulletin of Experimental Biology and Medicine* 2015; 159(3):380-383.
- [67] Michelle S. Antunes, André T.R. Goes, Silvana P. Boeira, Marina Prigol, Cristiano R. Jesse: Protective Effect Of Hesperidin In A Model Of Parkinson's Disease Induced By 6- Hydroxydopamine In Aged Mice. *Nutrition* 2014; 30(11-12):1415-22.
- [68] Lucian Hritcu, Harquin Simplicie Foyet, Marius Stefan, Marius Mihasan, Acha Emmanuel Asongalem, Pierre Kamtchoung: Neuroprotective Effect Of The Methanolic Extract Of Hibiscus Asp. *Journal of Ethnopharmacology* 2011; 137:585-591.
- [69] Young-Kyoung Ryu, Young Kang, Jun Go, Hye-Yeon Park, Jung-Ran Noh, Yong-Hoon Kim, Jung Hwan Hwang, Dong-Hee Choi, Sang-Seop Han, Won-Keun Oh, Chul-Ho Lee, And Kyoung-Shim Kim: Humulus Japonicus Prevents Dopaminergic Neuron Death In 6-Hydroxydopamine-Induced Models Of Parkinson's Disease. *J Med Food* 2017; 20(2):116-123.
- [70] Zahra Kiasalari, Tourandokht Baluchnejadmojarad, Mehrdad Roghani: Hypericum Perforatum Hydroalcoholic Extract Mitigates Motor Dysfunction and Is Neuroprotective in Intrastriatal 6-Hydroxydopamine Rat Model of Parkinson's Disease. *Cell Mol Neurobiol* 2016; 36(4):521-30.
- [71] Hannah Jang, Schwan Kim, Jae Man Lee, Yong-Seok Oh, Sang Myun Park and Sang Ryong Kim: Montelukast Treatment Protects Nigral Dopaminergic Neurons Against Microglial Activation in the 6-Hydroxydopamine Mouse Model of Parkinson's Disease. *Neuroreport* 2017; 28(5):242-249.

- [72] Bala V. Manyam, Muralikrishnan Dhanasekaran and Theodore A. Hare: Neuroprotective Effects of The Antiparkinson Drug Mucuna Pruriens. *Phytother. Res* 2004; 18:706–712.
- [73] Haiyan Lou, Xu Jing, Xinbing Wei, Huanying Shi, Dongmei Ren, Xiumei Zhang: Naringenin Protects Against 6-OHda-Induced Neurotoxicity Via Activation Of The Nrf2/Are Signaling Pathway. *Neuropharmacology* 2014; 79:380-8.
- [74] Musa V. Mabandla, Mpumelelo Nyoka, Willie M.U. Daniels: Early Use Of Oleanolic Acid Provides Protection Against 6-Hydroxydopamine Induced Dopamine Neurodegeneration. *Brain Research* 2015; 1622: 64–71.
- [75] Pallavi Shrivastava, Kumar Vaibhav, Rizwana Tabassum, Andleeb Khan, Tauheed Ishrat, Mohd. Moshahid Khan, Et Al: Anti-Apoptotic and Anti-Inflammatory Effect of Piperine on 6-OHda Induced Parkinson's Rat Model. *J Nutr Biochem* 2013; 24(4):680-7.
- [76] Nagaraja Haleagrahara, Cheng Jun Siew, Nilesh Kumar Mitra, Mangala Kumari: Neuroprotective Effect of Bioflavonoid Quercetin In 6-Hydroxydopamine-Induced Oxidative Stress Biomarkers In The Rat Striatum. *Neuroscience Letters* 2011; 500:139–143.
- [77] Saif Ahmad, M. Badruzzaman Khan, M. Nasrul Hoda, Kanchan Bhatia, Rizwanul Haque, Inayat Saleem Fazili, Arshad Jamal, Jafar Salamt Khan, Deepshikha Pande Katare: Neuroprotective Effect Of Sesame Seed Oil In 6-Hydroxydopamine Induced Neurotoxicity In Mice Model: Cellular, Biochemical And Neurochemical Evidence. *Neurochem Res* 2012; 37:516–526.
- [78] R. Vadivelan , N. Punitha, Triveni Jasti, Gautam Adhikari, T. Arun And Shem Joseph: Anti-Parkinson's Activity Of Sorafenib In 6-OHda Induced Rat Model. *Ijpsr* 2019; 10(9): 4257-4263.
- [79] M. H. Shalavadi, V. M. Chandrashekhar, S. P. Avinash, C. Sowmya, A. Ramkishan: Neuroprotective Activity of Stereospermum Suaveolens Dc Against 6-OHda Induced Parkinson's Disease Model. *Indian Journal of Pharmacology* 2012; 44(6):737-743.
- [80] Özge Güzelad, Ayşe Özkan, Hande Parlak, Osman Sinen, Ebru Afşar, Eren Ögüt, Fatoş Belgin Yıldırım, Mehmet Bülbül, Aysel Açar, Mutay Aslan: Protective Mechanism Of Syringic Acid In An Experimental Model Of Parkinson's Disease. *Metabolic Brain Disease* 2021; 36:1003–1014.
- [81] Saeideh Nourmohammadi, Sanaz Yousef, Mahboubeh Manouchehrabadi, Mona Farhadi, Zahra Azizi And Anahita Torkaman-Boutorabi: Thymol Protects Against 6-Hydroxydopamine-Induced Neurotoxicity In In Vivo And In Vitro Model Of Parkinson's Disease Via Inhibiting Oxidative Stress. *Bmc Complementary Medicine and Therapies* 2022; 40(22):1-14.
- [82] Reza Sedaghat, Mehrdad Roghani and Mohsen Khalili: Neuroprotective Effect of Thymoquinone, The Nigella Sativa Bioactive Compound, In 6-Hydroxydopamine-Induced Hemi-Parkinsonian Rat Model. *Iranian Journal of Pharmaceutical Research* 2014; 13(1): 227-234.
- [83] Jayasankar Kosaraju, Santhivardhan Chinni, Partha Deb Roy, Elango Kannan, A. Shanish Antony, M. N. Satish Kumar: Neuroprotective Effect Of Tinospora Cordifolia Ethanol Extract On 6-Hydroxy Dopamine Induced Parkinsonism. *Indian Journal of Pharmacology* 2014; 46(2):176-180.
- [84] Jie Ren, Ling Yuan, Wenbin Wang, Meiju Zhang, Qun Wang, Simin Li, Ling Zhang, Kun Hu: Tricetin Protects Against 6-OHda-Induced Neurotoxicity In Parkinson's Disease Model By Activating Nrf2/Ho-1 Signaling Pathway And Preventing Mitochondria-Dependent Apoptosis Pathway. *Toxicol Appl Pharmacol* 2019; 378:114617.
- [85] Tourandokht Baluchnejadmojarad, Nida Jamali-Raeufy, Sedigheh Zabihnejad, Nafiseh Rabiee, Mehrdad Roghani: Troxerutin Exerts Neuroprotection In 6-Hydroxydopamine Lesion Rat Model Of Parkinson's Disease: Possible Involvement Of Pi3k/Erβ Signaling. *Eur J Pharmacol* 2017; 801:72-78.
- [86] Rasha Abuthawabeh, Amjad N. Abu Irmaileh and Karem H. Alzoubi: The Beneficial Effect of Vanillin On 6-Hydroxydopamine Rat Model of Parkinson's Disease. *Restor Neurol Neurosci* 2020; 38(5):369-373.
- [87] Amit K. Sharma, Sparsh Gupta, Ranjan K. Patel And Neeta Wardhan: Haloperidol-Induced Parkinsonism Is Attenuated By Varenicline In Mice. *J Basic Clin Physiol Pharmacol* 2018; 29(4):395-401.
- [88] Angie M. Ameen, Amany Y Elkazaz, Hala M.F. Mohammad, Bassant M. Barakat: Anti-Inflammatory And Neuroprotective Activity Of Boswellic Acids In Rotenone Parkinsonian Rats. *Can J Physiol Pharmacol* 2017; 95(7):819-829.
- [89] Muhammad Aslam, Hammad Ahmed, Tayyaba Mumtaz And Gahzal Hakani: Antiparkinsonian Activity Of Aqueous Extract Of Agaricus Blazei Murill In Rotenone-Induced Parkinson's Disease. *Jpri* 2021; 33(33b):121-131.
- [90] Caner G'Unaydın, Bahattin Avci, Ayhan Bozkurt, Mehmet Emin Onger, Hakan Balci, S. Sirri Bilge: Effects Of Agomelatine In Rotenone-Induced Parkinson's Disease In Rats. *Neurosci Lett* 2019; 699:71-76.
- [91] Angie M. Ameen, Amany Y Elkazaz, Hala M.F. Mohammad, Bassant M. Baraka: Anti-Inflammatory and Neuroprotective Activity of Boswellic Acids in Rotenone Parkinsonian Rats. *Can J Physiol Pharmacol* 2017; 95(7):819-829.
- [92] Sawsan A. Zaitone, Eman Ahmed, Nehal M. Elsherbiny, Eman T. Mehanna, Mohammed K. El-Kherbetawy, Mohamed H. Elsayed, Duha M. Alshareef, Yasser M. Moustafa: Caffeic Acid Improves Locomotor Activity And Lessens Inflammatory Burden In A Mouse Model Of Rotenone-Induced Nigral Neurodegeneration: Relevance To Parkinson's Disease Therapy. *Pharmacol Rep* 2019; 71(1):32-41.
- [93] Poonam Thakur & Bimla Nehru: Inhibition of Neuroinflammation and Mitochondrial Dysfunctions by Carbenoxolone in The Rotenone Model of Parkinson's Disease. *Mol Neurobiol* 2015; 51(1):209-19.
- [94] Rania M. Salama, Ghada A. Abdel-Latif, Samah S. Abbas, Hekmat M. El Magdoub, Mona F. Schaalan: Neuroprotective Effect Of Crocin Against Rotenone-Induced Parkinson's Disease In Rats: Interplay Between Pi3k/Akt/Mtor Signaling Pathway And Enhanced Expression Of Mirna-7 And Mirna-221. *Neuropharmacology* 2020; 164:1-35.
- [95] Muthu Ramkumar, Srinivasagam Rajasankar, Veerapan Venkatesh Gobi, Udaiyappan Janakiraman, Thamilarasan Manivasagam, Arokiasamy Justin Thenmozhi, Musthafa Mohamed Essa, Ranganathan Chidambaram, Saravana Babu Chidambaram, Giles J. Guillemin: Demethoxycurcumin, A Natural Derivative Of Curcumin Abrogates Rotenone-Induced Dopamine Depletion And Motor Deficits By Its Antioxidative And Anti-Inflammatory Properties In Parkinsonian Rats. *Pharmacognosy Magazine* 2018; 14(53):9-16.
- [96] Mariama S. Azmy & Esther T. Menze & Reem N. El-Naga & Mariane G. Tadros: Neuroprotective Effects Of Filgrastim In Rotenone-Induced Parkinson's Disease In Rats: Insights Into Its Anti-Inflammatory, Neurotrophic, And Antiapoptotic Effects. *Mol Neurobiol* 2018; 55(8):6572-6588.
- [97] Shreesh Ojha, Hayate Javed, Sheikh Azimullah, Salema B. Abul Khair, M. Emdadul Haque: Glycyrrhizic Acid Attenuates Neuroinflammation And Oxidative Stress In Rotenone Model Of Parkinson's Disease. *Neurotox Res* 2016; 29(2):275-87.
- [98] Rosalba Siracusa, Maria Scuto, Roberta Fusco, Angela Trovato, Maria Laura Ontario, Roberto Crea, Rosanna Di Paola, Salvatore Cuzzocrea And Vittorio Calabrese: Anti-Inflammatory And Anti-Oxidant Activity Of Hidrox® In Rotenone-Induced Parkinson's Disease In Mice. *Antioxidants* 2020; 824(9):1-19.

- [99] Dharmendra Kumar Khatri & Archana Ramesh Juvekar: Propensity Of Hyoscyamus Niger Seeds Methanolic Extract To Allay Stereotaxically Rotenone-Induced Parkinson's Disease Symptoms In Rats. *Orient Pharm Exp Med* 2015; 1-13.
- [100] Rengasamy Balakrishnan, Dhanraj Vijayaraja, Thangavel Mohankumar, Dharmar Manimaran, Palanivel Ganesan, Dong-Kug Choi, Namasivayam Elangovan: Isolongifolene Mitigates Rotenone-Induced Dopamine Depletion And Motor Deficits Through Anti-Oxidative And Anti-Apoptotic Effects In A Rat Model Of Parkinson's Disease. *J Chem Neuroanat* 2021; 112:1-31.
- [101] Qiuyu Lin, Sen Hou, Yuyin Dai, Nan Jiang And Yingjie Lin: Monascin Exhibits Neuroprotective Effects In Rotenone Model Of Parkinson's Disease Via Antioxidation And Anti-Neuroinflammation. *Neuroreport* 2020; 31(9):637-643.
- [102] Hayate Javed, Sheikh Azimullah, Salema B. Abul Khair, Shreesh Ojha And M. Emdadul Haque: Neuroprotective Effect Of Nerolidol Against Neuroinflammation And Oxidative Stress Induced By Rotenone. *Javed Et Al. BMC Neurosci* 2016; 17(58):1-12.
- [103] Małgorzata Kujawska, Michael Jourdes, Monika Kurpiak, Michał Szulc, Hanna Szafer, Piotr Chmielarz, Grzegorz Kreiner, Violetta Krajka-Kuźniak, Przemysław Łukasz Mikołajczak, Pierre-Louis Teissedre and Jadwiga Jodynis-Liebert: Neuroprotective Effects of Pomegranate Juice against Parkinson's Disease and Presence of Ellagitannin-Derived Metabolite—Urolithin A—In the Brain. *Int. J. Mol. Sci* 2020; 21(202):1-20.
- [104] Marwa Y. Issa, Marwa I. Ezzat, Rabab H. Sayed, Eman M. Elbaz, Farghaly A. Omar, Engy Mohsen: Neuroprotective effects of pulicaria undulata essential oil in rotenone model of parkinson's disease in rats: Insights into its anti-inflammatory and anti-oxidant effects. *South African Journal of Botany* 2020; 132:289-298.
- [105] Haruka Kaji, Isao Matsui-Yuasa, Kayo Matsumoto, Ayano Omura, Kunio Kiyomoto, Akiko Kojima-Yuasa: Sesaminol Prevents Parkinson's Disease by Activating the Nrf2-Are Signaling Pathway. *Heliyon* 2020; 6(E05342):1-11.
- [106] Navneet Khurana, Asmita Gajbhiye: Ameliorative Effect of Sida Cordifolia in Rotenone Induced Oxidative Stress Model of Parkinson's Disease. *Neurotoxicology* 2013; 39:57-64.
- [107] Vaibhavi Peshattiwar, Suraj Muke, Aakruti Kaikini, Sneha Bagle, Vikas Dighe, Sadhana Sathaye: Mechanistic Evaluation Of Ursolic Acid Against Rotenone Induced Parkinson's Disease- Emphasizing The Role Of Mitochondrial Biogenesis. *Brain Res Bull* 2020; 160:150-161.
- [108] Neha Sharma, Navneet Khurana, Arunachalam Muthuraman, Puneet Utreja: Pharmacological Evaluation Of Vanillic Acid In Rotenone-Induced Parkinson's Disease Rat Model. *European Journal of Pharmacology* 2021; 903:1-13.
- [109] Neha Sharma and Bimla Nehru: Beneficial Effect of Vitamin E In Rotenone Induced Model of Pd: Behavioural, Neurochemical and Biochemical Study. *Exp Neurobiol* 2013; 22(3):214-223.
- [110] Sudhakar Pemminati, V Nair, Dorababu.P, Gopalakrishna Hn, Pai Mrs: Effect Of Aqueous Fruit Extract Of Emblica Officinalis On Haloperidol Induced Catalepsy In Albino Mice And. *Journal Of Clinical And Diagnostic Research* 2009; 3:1657-1662.
- [111] Chitra V, Manasa K, Mythili A, Tamilanban T, Gayathri K: Effect Of Hydroalcoholic Extract Of Achyranthes Aspera On Haloperidol-induced Parkinson's Disease In Wistar Rats. *Asian J Pharm Clin Res* 2017; 10(9):318-321.
- [112] Uzma Saleem, Zohaib Raza, Fareeha Anwar, Zunera Chaudary and Bashir Ahmad: Systems Pharmacology Based Approach to Investigate the In-Vivo Therapeutic Efficacy of Albizia Lebbeck (L.) In Experimental Model of Parkinson's Disease. *BMC Complementary and Alternative Medicine* 2019; 19(352):1-16.
- [113] Vandana S. Nade, Laxman A. Kawale, Shankar S. Zambre, Amit B. Kapure: Neuroprotective Potential Of Beta Vulgaris L. In Parkinson's Disease. *Indian Journal of Pharmacology* 2015; 47(4):403-408.
- [114] Uzma Saleem, Shabana Bibi, Muhammad Ajmal Shah, Bashir Ahmad, Ammara Saleem, Zunera Chauhdary, Fareeha Anwar, Nimra Javaid, Sundas Hira, Muhammad Furqan Akhtar, Ghulam Mujtaba Shah, Muhammad Saad Khan, Haji Muhammad, Muhammad Qasim, Mohammad Alqarni, Majed A. Algarni, Renald Blundell, Celia Vargas-De-La-Cruz, Oscar Herrera-Calderon, Reem Hasaballah Alhasani: Anti-Parkinson's Evaluation Of Brassica Juncea Leaf Extract And Underlying Mechanism Of Its Phytochemicals. *Frontiers in Bioscience-Landmark* 2021; 26(11):1031-1051.
- [115] Mohajjel Nayebi, Sheidaei H: Buspirone Improves Haloperidol-Induced Parkinson Disease in Mice Through 5-HT_{1A} Receptors. *Daru* 2010; 18(1):41-45.
- [116] Omar M.E. Abdel-Salam, Marawa El-Sayed El-Shamarka, Neveen A. Salem, Alaa El-Din M. Gaafar: Effects Of Cannabis Sativa Extract On Haloperidol-Induced Catalepsy And Oxidative Stress In The Mice. *Excli Journal* 2012; 11:45-58.
- [117] Uzma Saleem, Aisha Shehzad, Shahid Shah, Zohaib Raza, Muhammad Ajmal Shah, Shabana Bibi, Zunera Chauhdary, Bashir Ahmad: Antiparkinsonian Activity Of Cucurbita Pepo Seeds Along With Possible Underlying Mechanism. *Metab Brain Dis* 2021; 36(6):1231-1251.
- [118] Balvinder Kaur and Jyoti Saxena: Evaluation of Anti-Parkinson's Activity of Cyamopsis Tetragonoloba Methanol Plant Extract with Behavioral and Biochemical Analysis. *Ijpsr* 2021; 12(6):3236-3242.
- [119] Maham Sanawar, Uzma Saleem, Fareeha Anwar, Samra Nazir, Muhammad Furqan Akhtar, Bashir Ahmad & Tariq Ismail: Investigation Of Anti Parkinson Activity Of Dicyclomine. *Int J Neurosci* 2020; 1-14.
- [120] Harish G. Bagewadi, A. K. Afzal Khan: Evaluation of Anti-Parkinsonian Activity of Elaeocarpus Ganitrus on Haloperidol Induced Parkinson's Disease In Mice. *Int J Basic Clin Pharmacol* 2015; 4(1):102-106.
- [121] Sudhakar Pemminati, V Nair, Dorababu.P, Gopalakrishna Hn, Pai Mrs: Effect Of Aqueous Fruit Extract of Emblica Officinalis on Haloperidol Induced Catalepsy In Albino Mice And. *Journal of Clinical And Diagnostic Research* 2009; (3)1657-1662.
- [122] Ruby S and Jaykar B: Isolation of Pyran Composition and Anti-Parkinson's Activity of Euphorbia Cyathophora. *Asian Journal of Pharmaceutical Research and Development* 2019; 7(3):67-74.
- [123] Jitendra O. Bhangale and Sanjeev R. Acharya: Anti-Parkinson Activity of Petroleum Ether Extract of Ficus Religiosa (L.) Leaves. *Advances in Pharmacological Sciences* 2016; 9436106:1-9.
- [124] Werner J. Schmidt, Gertrud Schuster, Eva Wacker, Gabriela Pergande: Antiparkinsonian and Other Motor Effects of Flupirtine Alone and In Combination with Dopaminergic Drugs. *European Journal of Pharmacology* 1997; 327:1-9.
- [125] Kabra Mp, Bhandari Ss, Sharma A, Gupta Rb: Evaluation Of Anti-Parkinson's Activity Of Gentisic Acid In Different Animal Models. *Journal of Acute Disease* 2014; 141-144.
- [126] C. T. Chopde, M. S. Hote, S. N. Mandhane, And A. V. Muthal: Glucocorticoids Attenuate Haloperidol-induced Catalepsy Through Adrenal Catecholamines. *J Neural Transm [Gen Sect]* 1995; 102:47-54.

- [127]Graziella R. Molska, Lyvia Izaura G. Paula-Freire, Marna E. Sakalem, Daniele O. Köhn, Giuseppina Negri, Elisaldo A. Carlini & Fúlvio R. Mendes: Green Coffee Extract Attenuates Parkinson's-related Behaviors In Animal Models. *An Acad Bras Cienc* 2021; 93(4):1-21.
- [128]Awais Ali Zaidi, Mahtab Ahmad Khan, Zaib Ali Shahreyar And Hammad Ahmed: Lauric Acid:Its Role In Behavioral Modulation, Neuro-Inflammatory And Oxidative Stress Markers In Haloperidol Induced Parkinson's Disease. *Pak. J. Pharm. Sci* 2020; 33(2):755-763.
- [129]Halimah A. Adedeji, Ismail O. Ishola, Olufunmilayo O. Adeyemi: Novel Action Of Metformin In The Prevention Of Haloperidol-Induced Catalepsy In Mice: Potential In The Treatment Of Parkinson's Disease?. *Progress in Neuro-Psychopharmacology & Biological Psychiatry* 2014; 48:245–251.
- [130]Atul Kabra, Uttam Singh Baghel, Christophe Hano, Natalia Martins, Mohammad Khalid, Rohit Sharma: Neuroprotective Potential Of Myrica Esulenta In Haloperidol Induced Parkinson's Disease. *Journal of Ayurveda and Integrative Medicine* 2020; 11:448-454.
- [131]A S Rasheed S Venkataraman, K N Jayaveera, A Mohammed Fazil, K J Yasodha, M A Aleem, M Mohammed, Z Khaja, B Ushasri, H A Pradeep, M Ibrahim: Evaluation Of Toxicological And Antioxidant Potential Of Nardostachys Jatamansi In Reversing Haloperidol-Induced Catalepsy In Rats. *International Journal of General Medicine* 2010; 3:127-136.
- [132]S. Pemminati, V. Nair, P. Dorababu, H.N. Gopalakrishna, M.R.S.M. Pai: Effect of Ethanolic Leaf Extract of *Ocimum Sanctum* on Haloperidol-Induced Catalepsy in Albino Mice. *Indian J Pharmacol* 2007; 39(2):87-89.
- [133]Rashmi Anandpara, Bhakti Sojitra, Yash Nakhva, Tejas Ganatra: Evaluation And Comparision Of Anti Parkinson Activity Of Methanolic Extract Of *Phaseolus Vulgaris* With L-Dopa. *International Journal of Pharmacology Research* 2015; 5(2):103-108.
- [134]Santosh Kumar Vaidya, Dharmesh K. Golwala, Darpini S. Patel and Satyajit Sahoo: Evaluation of Antioxidant and Anti-Parkinson Activity of *Portulaca oleracea* Seed Methanolic Extract. *EJMP* 2020; 31(2):10-17.
- [135]Uzma Saleem, Zujajah Gull, Ammara Saleem, Muhammad Ajmal Shah, Muhammad Furqan Akhtar, Fareeha Anwar, Bashir Ahmad, Pharkphoom Panichayupakaranant: Appraisal Of Anti-Parkinson Activity Of Rhinacanthin-C In Haloperidol-Induced Parkinsonism In Mice: A Mechanistic Approach. *J Food Biochem* 2021; 45(E13677):1-13.
- [136]Uzma Saleem, Zunera Chauhdary, Zohaib Raza, Shahid Shah, Mahmood-Ur Rahman, Parwasha Zaib, And Bashir Ahmad: Anti-Parkinson's Activity Of *Tribulus Terrestris* Via Modulation Of Ache, A-Synuclein, Tnf-A, And Il-1 β . *Acs Omega* 2020; 5:25216–25227.
- [137]Prakash Chaudhary and Swati Dhande: Evaluation of Anti-Parkinson's Activity of Ethanolic Extract of *Tridax Procumbens* (Asteraceae). *Indian Journal of Natural Products and Resources* 2020; 11(1):9-17.
- [138]Amit K. Sharma, Sparsh Gupta, Ranjan K. Patel and Neeta Wardhan: Haloperidol-Induced Parkinsonism Is Attenuated By Varenicline In Mice. *J Basic Clin Physiol Pharmacol* 2018; 29(4):395-401.
- [139]Sushmita Singh, Imtiyaz Ansari: Evaluation of Antiparkinsonian Activity of Hydroalcoholic Extract of The Seeds Of *Vigna Aconitifolia* In Wistar Albino Rat. *Asian J Pharm Clin Res* 2019; 12(12):143-148.
- [140]Vinod Nair, Albina Arjuman, H. N. Gopalakrishna And M. Nandini: Effect Of *Withania Somnifera* Root Extract On Haloperidol-Induced Catalepsy In Albino Mice. *Phytother. Res* 2008; 22:243–246.
- [141]Humera Khatoon, Rahela Najam, Talat Mirza And Bushra Sikandar: Beneficial Anti-Parkinson Effects Of Camel Milk In Chlorpromazineinduced Animal Model: Behavioural And Histopathological Study. *Pak. J. Pharm. Sci* 2016; 29(5):1525-1529.
- [142]Sadaf Naeem, Rahila Najam, Saira Saeed Khan, Talat Mirza, Bushra Sikandar: Neuroprotective Effect of Diclofenac on Chlorpromazine Induced Catalepsy In Rats. *Metab Brain Dis* 2019; 34(4):1191-1199.
- [143]Rashmi Anandpara, Bhakti Sojitra, Yash Nakhva, Tejas Ganatra: Evaluation And Comparision Of Anti Parkinson Activity Of Methanolic Extract Of *Phaseolus Vulgaris* With L-Dopa. *International Journal of Pharmacology Research* 2015; 5(2):103-108.



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