



# IJRASET

International Journal For Research in  
Applied Science and Engineering Technology



---

# INTERNATIONAL JOURNAL FOR RESEARCH

IN APPLIED SCIENCE & ENGINEERING TECHNOLOGY

---

**Volume:** 11    **Issue:** III    **Month of publication:** March 2023

**DOI:** <https://doi.org/10.22214/ijraset.2023.49559>

[www.ijraset.com](http://www.ijraset.com)

Call:  08813907089

E-mail ID: [ijraset@gmail.com](mailto:ijraset@gmail.com)

# Total Leaf Protein Content: A Measure to Study the Phytotoxic Effects of Sulfur Dioxide on Crop Plants

Aprajita Chauhan

Department of Chemistry Sri Aurobindo College (University of Delhi), Malviya Nagar, New Delhi-110017, INDIA

**Abstract:** Effect of different concentrations on total leaf protein content was studied in three economically important plant species, viz., tomato, mung bean and maize. Different treatments of SO<sub>2</sub> exposure were administered in artificial fumigation chambers. Maize showed least reduction in total proteins. Tomato exhibited maximum decline in protein content after SO<sub>2</sub> fumigation. Relationship between individual and interactive effects of SO<sub>2</sub> concentration were analyzed with the help of a statistical regression model.

Present study helps to establish a correlation between the total leaf protein content, free radicals, activities of antioxidant enzymes like superoxide dismutases and peroxidases and plant sensitivity to SO<sub>2</sub> under ambient conditions as well as in greenhouse environment.

**Keywords:** controlled SO<sub>2</sub>-fumigation, total leaf protein content, tomato, mung bean, maize.

## I. INTRODUCTION

Sulfur dioxide is one of the dangerous phytotoxicants in ambient as well as under greenhouse conditions. Exposure to low concentrations of SO<sub>2</sub> is known to induce several physiological and biochemical modifications in plants, as documented and reviewed by many investigators like Chauhan, (1989a), Darall, (1989), Varshney et al., (2009), Rai et al., (2011), Singh et al., (2012), Brahmachari and Kundu, (2017), and Lee et al., (2017). Total leaf protein content constitutes one of the important parameters for healthy plant metabolism. Investigators like Agrawal et al., (2006), Varshney et al., (2009), Singh et al., (2012), and Lee et al., (2017) have observed a decrease in total proteins in SO<sub>2</sub>-fumigated plants in many plants. Alohisen (2014) attributes this decline primarily to the free-radical induced breakdown of existing proteins, resulting in an increase in the total free amino acid content. Reduced *de novo* synthesis of proteins is another reason for the decrease in protein content. The relationship between total protein content and the activity of antioxidant enzymes like peroxidases and superoxide dismutase (SOD) helps to explain the possible mechanisms of SO<sub>2</sub>-tolerance by many plants. Present investigation on three economically important crop plants explains the levels of total leaf proteins in relation to SO<sub>2</sub> stress. The combined effects of SO<sub>2</sub> dosage and exposure time have been analysed using a regression model.

## II. MATERIAL AND METHODS

### A. Plant Material and Fumigation System

Three economically important cultivated plant species viz., *Vigna radiata* (L.) Wilczek [Mung bean], *Solanum esculentum* (= *Lycopersicon esculentum* Mill.) [Tomato], and *Zea mays* L. [Maize] were grown from seeds in the nursery. Fifteen-day-old seedlings of these plants were subjected to different SO<sub>2</sub> treatments through an artificial fumigation system. Sulfur dioxide was generated from an aqueous solution of Na<sub>2</sub>S<sub>2</sub>O<sub>5</sub> and circulated over the plants kept in specially designed closed-top fumigation chambers (1 x 1 x 1m = 1m<sup>3</sup>) at temperatures ranging between 25-29°C ± 1°C and at a Relative humidity (RH) of 60 ± 5%. As a source of illumination, two metal halide lamps of 200W each were employed following a light/dark cycle of 12/12 hours.

### B. Fumigation Protocol

Artificial fumigations were carried out according to the following protocols detailed by Chauhan, (1989a):

Treatments: T-1 = 0.05 ppm (134.0 μg m<sup>-3</sup> SO<sub>2</sub>) [x 4h], T-2 = 0.1 ppm (268.0 μg m<sup>-3</sup> SO<sub>2</sub>) [x 2h], and T-3 = 0.2 ppm (536.0 μg m<sup>-3</sup> SO<sub>2</sub>) [x 1h] for 60 days. Thus, the SO<sub>2</sub> dose was kept constant. Among the above, *V. radiata* was fumigated for only 45 days. Controls (C) were maintained simultaneously by exposing the plants to air alone. Fresh leaf tissue (0.2g) was homogenized in 10 ml of 5% TCA and total protein content was estimated following the method of Lowry et al., (1951).

### III. RESULTS AND DISCUSSION

Leaves of *Vigna radiata* showed considerable reduction in total leaf proteins following SO<sub>2</sub> treatment. A progressive reduction in protein content was observed with respect to the dosage, viz., 39.34, 22.10 and 16.57 per cent for treatments T-3, T-2, and T-1 respectively for 45-day fumigation (Table 1, Fig. 1). Whereas the total leaf protein content in control plants of *Solanum esculentum* (= *Lycopersicon esculentum*) was slightly less than that recorded for *Vigna*, there was much pronounced reduction in leaf proteins in all the three treatments. The reduction values were much higher in T-1, T-2 and T-3 over the corresponding values in *Vigna*. For instance, values for total leaf protein content in the treatments T-1, T-2, and T-3 for 60 days were 30.21, 57.94, and 62.68% respectively (Table 2, Fig. 2), which were considerably higher than the other plant species investigated. SO<sub>2</sub>-fumigated plants of *Zea mays* exhibited least reduction in total leaf protein content. 60-days fumigation cycle with T-1, T-2, and T-3 treatments resulted in a reduction in protein content of only 14.14, 18.75, and 21.87% respectively (Table 3, Fig. 3).

Investigators like Malhotra and Khan (1984), Hamid and Jawaid, (2009), Rai et al., (2011), Singh et al., (2012), Brahmachari and Kundu, (2017), and Lee et al., (2017) have reported a general decrease in protein content in SO<sub>2</sub>-fumigated plants in several cultivated and wild species. Reduction in protein content can be attributed to a) enhanced sulphitolysis, b) free radical reactions, and c) reduction in *de novo* protein synthesis, as suggested by Lenzian and Unsworth, (1983). Production of HSO<sub>3</sub><sup>-</sup> and SO<sub>3</sub><sup>-</sup> ions induces cleavage of disulphide linkages in proteins giving thiols and sulphonates. According to Shimazaki et al., (1980) sulphitolysis of the disulphide bond causes unfolding of protein molecules. Free radicals generated during SO<sub>2</sub> oxidation to SO<sub>4</sub><sup>2-</sup>, may induce lipid peroxidation, which constitutes an important mechanism of membrane deterioration in SO<sub>2</sub>-fumigated plants, as has been shown by Irigoyen et al., (1992). In addition, SO<sub>2</sub>-induced free radicals also inactivate proteins by modifying their amino acid residues (Wolff et al., 1986), thereby rendering many vital enzymes inactive. A distinct relationship seems to have been established regarding the effects of SO<sub>2</sub> on total proteins and the concentration of antioxidant enzymes. Peroxidases and SOD have been shown to play a vital role in SO<sub>2</sub> tolerance, as reported by Chauhan, (1989b), Singh et al., (2012), Brahmachari and Kundu, (2017), and Lee et al., (2017). *Vigna* and *Zea mays*, which show high SOD and peroxidase content, recorded less lipid peroxidation (Chauhan 1989b, 2015). Reduction in total leaf protein content after SO<sub>2</sub> exposure was also less in these two species as compared to tomato, which incidentally had low activities of these two enzymes. In addition, the emission of volatiles like ethylene, ethane, acetaldehyde and ethanol from SO<sub>2</sub>-treated plant tissues increase dramatically prior to the appearance of visible injury symptoms. Chauhan, (1990) reported that the spurt in volatile emissions is indicative of lipid peroxidation due to sulphate oxidation-induced free radical generation.

Interestingly, recent investigations, as reviewed by Li et al., (2022) also tend to implicate SO<sub>2</sub> as a vital signalling molecule. SO<sub>2</sub> fumigation is now thought to reprogramme the expression of many genes thereby modulating physiological processes like seed germination, stomatal action, fruit-keeping and response to environmental stress.

**TABLE 1. EFFECT OF SO<sub>2</sub> TREATMENTS ON TOTAL LEAF PROTEINS IN *V. radiata***

Period of Fumigation (Days)	15		30		45	
TREATMENT Conc. (ppm) Time (h)	Total Leaf Protein Content (mg/g f wt.)	Percent Reduction	Total Leaf Protein Content (mg/g f wt.)	Percent Reduction	Total Leaf Protein Content (mg/g f wt.)	Percent Reduction
C-1 (6×4)	9.42±0.40		11.73±0.754		14.06±1.00	
T-1 (0.05×4)	8.711±1.14	7.5	10.08±0.4012	14.06	11.73±0.686	16.57
C-2 (6×2)	7.15±0.547		9.2±0.628		10.95±0.374	
T-2 (0.10×2)	6.26±0.282	12.56	7.8±0.249	15.20	8.53±1.13	22.10
C-3 (6×1)	7.37±0.426		9.6±1.13		15.02±0.756	
T-3 (0.20×1)	6.04±0.764	18.04	7.68±0.453	20.80	9.11±1.457	39.34

Mean (SD) of 5 replicates C-1, C-2, C-3 : Controls [air time (h)];  
T-1, T-2, T-3 : Treatments [Conc. of SO<sub>2</sub> (ppm) Exposure time (h)]

**SIGNIFICANCE OF FACTORIAL EFFECTS : *V. radiata***

Period of fumigation (Days)	15				30				45			
	df	Sum of Squares	Mean Source	F	df	Sum of Squares	Mean Sources	F	df	Sum of Squares	Mean Sources	F
SO <sub>2</sub> Conc. (ppm)	17	3022.50	77.30	2.34**	17	4620.89	42.9	1.51*	17	7325.4	96.3	1.09*
Exposure Time (h)	27	2964.50	19.40	0.60	27	4629.54	51.64	1.81**	27	7399.83	170.67	1.94**
SO <sub>2</sub> Conc. Exposure Time	53	3074.96	129.76	3.93**	53	4700.2	123.0	4.32*	53	7583.90	354.74	4.04*
Error	10		32.96		10		28.46		10		87.7	

Levels of significance : \*\* P < 0.1 ; \* P < 0.25

**TABLE 2. EFFECT OF SO<sub>2</sub> TREATMENTS ON TOTAL LEAF PROTEINS IN *S. esculentum***

Period of Fumigation (Days)	15		30		45		60		
	Treatment Conc. (ppm) Time (h)	Total Leaf Protein Content (mg/g f wt.)	Percent Reduction	Total Leaf Protein Content (mg/g f wt.)	Percent Reduction	Total Leaf Protein Content (mg/g f wt.)	Percent Reduction	Total Leaf Protein Content (mg/g f wt.)	Percent: Reduction
C-1 (0x4) T-1 (0.05x4)		7.4±0.432		9.28±0.730		11.33±1.247		11.75±1.224	
		6.73±0.066	9.0	7.17±0.733	22.73	8.6±0.588	24.09	8.82±1.290	30.21
C-2 (0x2) T-2 (0.10x2)		6.40±0.821		7.82±1.28		9.46±0.653		9.51±1.188	
		5.57±0.871	12.96	5.57±0.475	28.77	4.22±0.553	55.57	4.02±1.474	57.94
C-3 (0x1) T-3 (0.20x1)		6.4±0.821		7.82±1.28		9.46±0.653		9.51±1.589	
		5.08±0.472	20.63	5.24±0.691	33.00	3.76±0.4618	59.84	3.55±0.2948	62.68

Mean (± SD) of 5 replicates C-1, C-2, C-3 : Controls [air × time (h)];

T-1, T-2, T-3 : Treatments [Conc. of SO<sub>2</sub> (ppm) × Exposure time (h)];

**SIGNIFICANCE OF FACTORIAL EFFECTS : *S. esculentum***

Period of fumigation (Days)	15				30				45				60			
	df	Sum of Squares	Mean Source	F	df	Sum of Squares	Mean Sources	F	df	Sum of Squares	Mean Sources	F	df	Sum of Squares	Mean Sources	F
SO <sub>2</sub> Conc. (ppm)	17	2018.38	9.60	0.43	17	2796.52	31.62	0.69*	17	3415.33	126.85	2.4**	17	3598.32	145.52	1.93
Exposure Time (h)	26	2033.19	21.40	0.96	26	2837.01	72.11	1.58**	26	3570.47	281.99	5.34**	26	3729.33	276.53	3.66**
SO <sub>2</sub> Conc. Exposure Time	53	2064.96	53.18	2.38	53	2914.28	149.38	3.27**	53	3750.12	461.64	8.74**	53	3950.28	497.48	6.59**
Error	10		22.17		10		45.65		10		52.8		10		75.43	

**TABLE 3. EFFECT OF SO<sub>2</sub> TREATMENTS ON TOTAL LEAF PROTEINS IN *Z. mays***

Period of Fumigation (Days)	15		30		45		60	
TREATMENT Conc. (ppm) Time (h)	Total Leaf Protein Content (mg/g f wt.)	Percent Reduction	Total Leaf Protein Content (mg/g f wt.)	Percent Reduction	Total Leaf Protein Content (mg/g f wt.)	Percent Reduction	Total Leaf Protein Content (mg/g f wt.)	Percent Reduction
C-1 (0×4)	7.84±0.1666		14.86±0.914		17.04±0.514		19.8±0.242	
T-1 (0.05×4)	7.55±0.2962	3.6	13.60±0.4618	8.4	15.71±0.412	7.89	16.7±0.50	14.14
C-2 (0×2)	6.0±1.2247		11.55±1.0		12.11±1.09		16.0±0.92	
T-2 (0.10×2)	5.77±1.238	3.83	10.26±1.52	11.6	10.33±0.422	14.69	13.0±0.845	18.75
C-3 (0×1)	8.13±0.4000		15.0±1.33		16.38±1.94		14.22±2.19	
T-3 (0.20×1)	7.75±0.8931	4.32	13.28±1.19	11.46	13.25±0.390	19.10	11.11±0.993	21.87

Mean (± SO) of 5 replicates C-1, C-2, C-3 : Controls [air × time (h)]

T-1, T-2, T-3 : Treatments [Conc. of SO<sub>2</sub> (ppm) × Exposure time (h)]

**SIGNIFICANCE OF FACTORIAL EFFECTS : *Z. mays***

Period of fumigation (Days)	15				30				45				60			
Source of Variation	df	Sum of Squares	Mean Source	F	df	Sum of Squares	Mean Sources	F	df	Sum of Squares	Mean Sources	F	df	Sum of Squares	Mean Sources	F
SO <sub>2</sub> Conc. (ppm)	17	2835.0	52.9	2.05*	17	9398.15	126.0	2.06**	17	10665.18	251.30	1.070*	17	12656.90	296.00	2.18**
Exposure Time (h)	26	2783.3	1.2	0.04	26	9300.60	28.0	0.50	26	10504.37	91.0	0.38	26	12488.48	127.58	0.94*
SO <sub>2</sub> Conc. X Exposure Time	53	2861.6	79.5	3.12**	53	9487.56	215.96	3.54**	53	10990.69	576.8	2.46	53	12919.60	558.70	4.12
Error	10		25.4		10		61.0		10		234.8		10		135.52	

Levels of significance : \*\* P < 0.50 ; \* P < 0.25 – 0.1

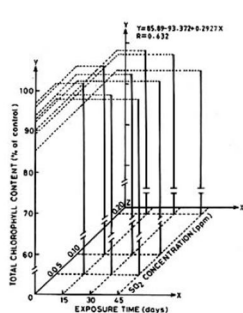


Fig.1. *Vradiata*

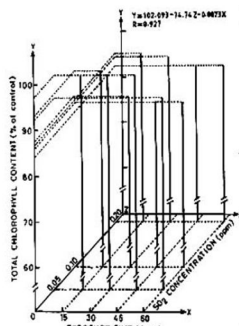


Fig.2. *S. esculentum*

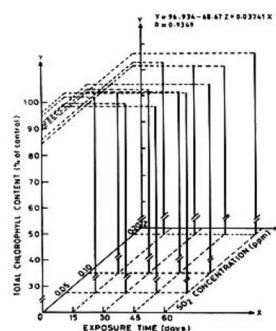


Fig.3. *Z. mays*

**Figs 1-3. Significance of Factorial Effects**

#### IV. ACKNOWLEDGEMENT

I am grateful to Prof. C.K.Varshney for his guidance at the School of Environmental Sciences, Jawaharlal Nehru University, New Delhi, India.

#### REFERENCES

- [1] M. Agrawal, B. Singh, S.B. Agrawal, J.N.B. Bell, and F.Marshall , "The effect of air pollution on yield and quality of mungbean grown in periurban areas of Varanasi," *Water, Air, Soil Poll.*, vol. 169, pp. 239-254, 2006.
- [2] A.A.Almohisen,"Response of free amino acids in four legumes plans to air pollution," *J.Biol. and Today'sWorld.*, vol. 3(8), pp.169-173, 2014.
- [3] S. Brahmachari and S. Kundu,"SO<sub>2</sub> stress: Its effect on plants, plant defence responses and strategies for developing enduring resistance. *Int. Adv. J. Sci. Eng. and Technol.*, vol. 4(7), pp.303- 309, 2017.
- [4] A. Chauhan ,1989a. Effect of Sulphur Dioxide on Plants at Biochemical and Physiological Levels. PhD. Thesis, University of Garhwal , India.
- [5] A. Chauhan, "Superoxide dismutase-a bioindicator of plant response to SO<sub>2</sub>-stress," in *Proc. Int. Sym. Pl. and Poll. in Dev. and Develop. Coun.*, Izmir, Turkey, p.549-568.
- [6] A. Chauhan, "Early diagnosis of SO<sub>2</sub>-stress by volatile emissions in some crop plants," *Oecologia*, vol. 84, pp. 289-294, 1990.
- [7] A. Chauhan, "Peroxidase activity as an indicator of SO<sub>2</sub> -tolerance in crop plants," *Int. Jour. Innov. Res. and Dev.*, vol.4 (13), pp. 45-50, 2015.
- [8] N.M Darall, "The effect of air pollutants on physiological processes in plants," *Plant Cell Environ.*, vol. 12, pp. 1-30, 1989.
- [9] N.Hamid and F. Jawaid, "Effect of short-term exposure of two different concentrations of sulphur dioxide and nitrogen mixture on some biochemical parameter of soybean (*Glycine max (L.) Merr.*," *Pak. J. Bot.*, vol.41(5), pp.2223-2228, 2009.
- [10] J.I. Irigoyen , D.W. Emerich , and M. Sanchez-Diaz, "Alfalfa leaf senescence induced by drought stress: photosynthesis, hydrogen peroxide metabolism, lipid peroxidation and ethylene evolution," *Physiol. Plant.*, vol. 84, pp. 67-72, 199
- [11] H. K. Lee, I. Khaine, M. J. Kwak, J.H. Jang, T.Y.Lee, J.K.Lee, L.R.Kim , W.I.Kim, K.S.Oh, and S.Y.Woo, "The relationship between SO<sub>2</sub> exposure and plant physiology: A mini review," *Hortic. Environ. Biotechnol.*, vol. 5(6), pp. 523-529, 2017.
- [12] K.J.Lendzian and MH. Unsworth, *Ecophysiological Effects of Atmospheric Pollutants*, O.L. Lange, P.S.Nobel , C.B.Osmond , and H.Ziegler, Eds. Berlin, Germany: Springer-Verlag, 1983.
- [13] Z.H. Li, X.E. Li, and H.Y. Chen, " Sulfur dioxide: An emerging signaling molecule in plants." *Front. Pl. Sci.*, vol. 13 , pp. 891626, 2022.
- [14] O.H. Lowry, N.J. Rosenbrough, A.L. Farr, and R.J. Randall, "Protein measurement with the folin phenol reagent," *J. Biol. Chem.* vol.193, pp. 267-275, 1951.
- [15] S.S.Malhotra and A.A.Khan, *Biochemical and Physiological Impact of Major Pollutants*. In M. Treshow, Ed. Chichester, John Wiley & Sons, pp. 1984.
- [16] R. Rai , M. Rajput, M. Agrawal , and S.B. Agrawal, "Gaseous air pollutants: a review on current and future trends of emissions and impact on agriculture" *J. Sci. Res. B.H.U.*, vol. 55, pp. 77-102, 2011.
- [17] K. Shimazaki , T. Sakaki, and K. Sugahara. "Active oxygen participation in chlorophyll destruction and lipid peroxidation in SO<sub>2</sub>- fumigated leaves of spinach," *Res. Rep. Nat. Inst. Environ. Sci. Ibarki, Japan*, vol. 11, pp. 91-101, 1980.
- [18] L.P.Singh , S.S.Gill , R. Gill, and N Tuteja, *Mechanism of Sulphur Dioxide Toxicity and Tolerance in Crop Plants*, N.Tuteja, A.F.Tiburcio, SS.Gill, and R.Tuteja, Eds. Weinheim, Germany: Wiley-VCH Verlag GmbH & KGaA, 2012.
- [19] C.K.Varshney, C.K. Garg, W.K. Lauenroth, and R.K.Heitschmidt, "Plant responses to sulphur dioxide pollution," *CRC Crit. Rev. Environ. Cont.*, vol. 9, pp. 27- 49, 2009.
- [20] S.P.Wolff , A. Garner, and R.T. Dean, "Free radicals, lipids and protein degradation." *Trends Biochem. Sci.*,vol 11, pp. 27-31, 1986.



10.22214/IJRASET



45.98



IMPACT FACTOR:  
7.129



IMPACT FACTOR:  
7.429



# INTERNATIONAL JOURNAL FOR RESEARCH

IN APPLIED SCIENCE & ENGINEERING TECHNOLOGY

Call : 08813907089  (24\*7 Support on Whatsapp)