



IJRASET

International Journal For Research in
Applied Science and Engineering Technology



INTERNATIONAL JOURNAL FOR RESEARCH

IN APPLIED SCIENCE & ENGINEERING TECHNOLOGY

Volume: 7 Issue: VII Month of publication: July 2019

DOI: <http://doi.org/10.22214/ijraset.2019.7062>

www.ijraset.com

Call:  08813907089

E-mail ID: ijraset@gmail.com

The Exogenous Effect of Gibberillic Acid on Enzyme Activities in Salt Stressed Rice Cultivars during Seed Germination

Dr. K. Krishna.

Associate Professor, P.G. Department of Botany, Yuvaraja's College, Constituent autonomous college, University of Mysore, Mysore, Karnataka, India.

Abstract: The amino acid content was increased under GA_3 treatment and also under combination of GA_3 and NaCl in both the cultivar. In salt sensitive Jyothi cultivar more protein was breakdown to amino acid hence the amino acid content increased, in salinity BPT Super cultivar the amino acid content decreased. It seems to be more tolerant when compared to the Jyothi cultivar. Therefore GA_3 showed the salinity mitigation role.

α - and β -amylase activity was found to be decreased under salinity and the increase was observed under GA_3 application. Combination of GA_3 and NaCl also increased the α - and β -amylase activity, the least affected salinity in BPT Super. In Jyothi cultivar salinity was more affected. Catalase activity was increased under salinity in both the cultivar. Salinity was increased under GA_3 treatment and also under the combination of GA_3 and NaCl, so catalase activity was least affected due to the salinity in both the paddy cultivars. With respect to all the parameters Jyothi paddy cultivar is more salt sensitive when compared to the BPT Super paddy cultivar. BPT Super shows better salinity tolerance when compared to the Jyothi cultivar and salinity alleviating role of GA_3 was observed.

Keywords: Rice, NaCl, GA_3 , Protein, Amino acids, α and β amylase and catalase.

I. INTRODUCTION

Rice (*Oryza sativa* L.) is a salt sensitive monocot and widely grown crop in tropical and subtropical regions. It is one the main staple foods for nearly two third of the population of the World (Roy *et al.*, 2012), high rice consumption, degradation of soil and water quality around globe have focused urgent attention to understand the response of this important crop towards abiotic stresses.

Salinity is estimated that over 800 million hectares of land in the World are affected (Munns, 2005; Kumar *et al.*, 2010 and Tavakkoli *et al.*, 2011). Salt stress in soil is one of the major stresses especially in arid and semi arid regions and can severely limit plant growth and productivity by reducing osmotic potential, ion toxicity creation, uptake disarrangement, ion imbalance and can cause disorders in enzyme activities and metabolic activities in plants (Hasegawa *et al.*, 2000; Basu *et al.*, 2002; Murphy *et al.*, 2003; Islam *et al.*, 2008). These activities could affect morphological parameters and will reduce vegetative growth (Linghe and Shannon, 2000; Rogers *et al.*, 2004) and chlorosis is common morphological and physiological characteristic in response to salt stress (Harinasut *et al.*, 2000). Chlorophyll pigment is a good indicator of plant nutrient stress during growing period and content of chlorophyll in the plant leaves indicates the growth of the crops. Synthesis and integrity of chlorophyll level may vary due to salt stress (Santo, 2004; Rout *et al.*, 1997). Chlorophyll content of salt stressed rice can be described as functions of the sodium content (Yeo and Flowers, 1983). Excess of NaCl to plants involves changes in their morphology, physiology and metabolism (Hilal *et al.*, 1998; Rahman *et al.*, 2008) and consequently reducing plant dry weight (Zeng and Shannon, 2000; Roger *et al.*, 2009) and dry matter production (Mansour and Salam, 2007) and ultimately crop yield (Jamil *et al.*, 2010; Osakabe *et al.*, 2011).

Several strategies have been proposed to alleviate the degree of cellular damage caused by abiotic stress and to improve crop salt tolerance. Among them exogenous application of compatible osmolytes such as proline, glycine betaine, trehalose, auxin, gibberellins, (Kim *et al.*, 2006; Misratia *et al.*, 2013; Rahdari and Hoseini, 2015), Methyl Jasmonate, Sucrose (Siringam *et al.*, 2012), Spermidine (Saleethong *et al.*, 2016) had gained considerable attention in mitigating the effect of salt stress. Plant growth regulators and other groups of chemicals have been used to treat rice plants exogenously at various growth stages to increase salt tolerance by alleviating salt induced damages and lead to improved growth and productivity (Roychoudhury *et al.*, 2011; Plaut *et al.*, 2013).

A few studies have however, demonstrated the ability of GA₃ to overcome adverse effects of NaCl stress (Chakraborti and Mukherji, 2003; Misratia *et al.*, 2013) GA₃ basically stimulated the inter node elongation, stem elongation and also control the various aspects of seed germination, mobilization of reserves, floral initiation, sex determination and fruit set. The present work was conducted to examine whether NaCl inhibited seed germination, growth and synthesis of chlorophyll content and to determine the influence of GA₃ when exposed to NaCl stress.

II. MATERIALS AND METHODS

Paddy seed sample of cultivars Jyothi and BPT super were procured from VC Farm, Regional Agricultural Research Station, University of Agriculture science, Mandya, Karnataka. Seeds of uniform size were selected and surface sterilized using 0.01% mercuric chloride (HgCl₂) for two minutes. The seeds were washed thoroughly with distilled water for several times and soaked for 24 hours in distilled water (control) and different concentrations of GA₃ i.e 100ppm and 200ppm and in NaCl i.e. 200mM and 300mM were used to soak the seeds as well as combination of NaCl and GA₃ seeds were also soaked. The germination studies were carried out as per International Seed Testing Association (2009). Five sets of each concentration of GA₃, NaCl and combination of both were maintained along with control. The seeds allowed to germinate for 14days as per ISTA, 2009 and then analyse for morphological and biochemical parameters.

Estimation of total protein content by Lowry *et al.*, (1951) had expressed in µg protein/ml and total amino acids contents by Moore and Stein (1948). α and β amylase activity was calculated as per Bernfeld (1955) and The enzyme activity is expressed as µmoles of glucose released/gm. Catalase activity was assayed according to Luck (1974).

III. RESULTS AND DISCUSSION

A. Protein

The test was conducted to study the effect of salinity on two rice cultivars and effect of GA₃ on Jyothi and BPT Super cultivar. Both the cultivar showed decrease in the protein content under salinity and the increase in protein content was observed under combination of NaCl and GA₃. The protein content was most affected in Jyothi than the BPT Super cultivar under salinity condition.

B. Effects Of Salinity On Protein Content

Protein content decreased in most of the plant species under Sodium chloride stress (Misra, *et al.*, 1997). A continuous decrease in protein content with increase in salt stress (Sunita Danai-Tambhale, 2011). The reason for decrease of protein content under salinity was breakdown of surface protein and production of free amino acid (Parvaneh rahdari, *et al.*, 2015).

C. Effects of GA₃ on Protein Content

Gibberellin treatment can lead to increased levels of protein, during the stress condition there will be increase in the nitrate reductase enzyme activity in the protein surface tension, and increase in the protein content (Masroor, *et al.*, 2006).

D. Amino Acid

In the present study the total amino acid content of two rice cultivar was estimated under salinity, total amino acid content of BPT Super was decreased but the total amino acid content of Jyothi cultivar was increased under salinity and under GA₃ treatment and combination of NaCl and GA₃ treatment in both the cultivars there was an increase in total amino acid content was observed.

E. Effects Of Salinity On Amino Acid Content

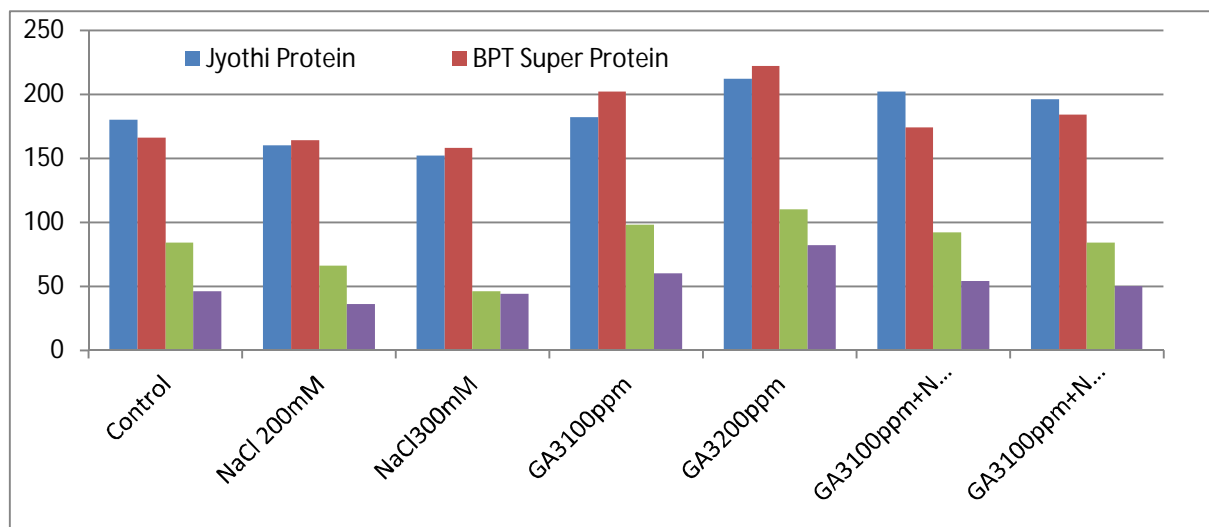
Total amino acid content was increased under salinity treatment and GA₃ treatment GA₃ treatment lead to the qualitative and quantitative changes in free amino acid, hence the increase in total amino acid content was observed in seeds treated with GA₃. Under salinity increase in amino acid was caused by proteolysis a great accumulation of protein amino acids, and disturbance of nitrogen metabolisms, and accumulation of non protein amino acids or ammonia was the reason for increase in total amino acid content under salinity (Yamamoto, *et al.*, 2011).

In general due to the salinity surface breakdown and production of free amino acids led to the increase in amino acid (Parvaneh rahdari, *et al.*, 2015). Total amino acid level in tolerant variety decreased under saline condition as reported by Saikat Paul, *et al.*, (2016). Total amino acids were also increased under salt stress as reported by Yamamoto, *et al.*, (2011).

Table 1: Effect of different concentrations of NaCl, GA₃ and combination of NaCl and GA₃ on Protein and amino acids content (µg/ml) of Jyothi and BPT Super paddy cultivars on 14th day.

Cultivar	Parameters	Control	NaCl 200mM	NaCl 300mM	GA ₃ 100ppm	GA ₃ 200ppm	GA ₃ 100ppm + NaCl 200mM	GA ₃ 200ppm + NaCl 300mM
Jyothi	Protein	180 ^c	160 ^d	152 ^e	182 ^e	212 ^a	202 ^b	196 ^b
	Amino acids	84 ^c	66 ^d	46 ^e	98 ^b	110 ^a	92 ^b	84 ^c
BPT Super	Protein	166 ^e	164 ^e	158 ^f	202 ^b	222 ^a	174 ^d	184 ^c
	Amino acids	46 ^d	36 ^e	44 ^d	60 ^b	82 ^a	54 ^{bc}	50 ^{cd}

Means followed by the same letter within a row are not significantly different as indicated by Scheffe ($P \leq 0.05$). Significant at $P \leq 0.001$.



Graph 1: Effect of different concentrations of NaCl, GA₃ and combination of NaCl and GA₃ on protein and amino acids content (µg/ml) of Jyothi and BPT Super paddy cultivars on 14th day.

F. α -Amylase Activity

Starch is the most abundant reserve food material in rice seeds. α -amylase is a crucial enzyme that participates in the degradation of starch granules into small organic molecules to provide energy and nutrients for seed germination. The study was done on the effect of salinity on α -amylase activity of rice seed where it was inhibited. Application of GA₃ on two rice cultivars there was an increase in α -amylase content. In the GA₃ 100ppm+NaCl 200mM quantitative data demonstrated that α -amylase activity was significantly increased compare to control.

G. Effect of Salinity on α -Amylase Activity

Seed germination is dependent on the degradation of storage reserves in mature seeds and the sugars from starch hydrolysis are the major source of energy for seedling emergence (Beck and Ziegler, 1989). α -amylase is the major enzyme involved in starch mobilization thus α -amylase activity is an important factor in seed germination. In this study quantitative data demonstrated that NaCl induced the inhibition of activity of α -amylase due to enhancing bioactive GA inactivation.

The reduction in α -amylase activity under salinity could account for the reduction in concentration and translocation of free sugars into the embryo axes during germination and early growth Saikat Paul., *et al.*, (2016) and due to the less imbibitions effect, because water as a solvent is necessary to stimulate the activity of this enzyme. Sodium chloride reduced α -amylase activity in germinating rice seeds to varying degrees even at low NaCl concentration (Shereen *et al.*, 2011). Salinity inhibits rice seed germination by decreasing the bioactive GA₃ content, as a result of an increase in bioactive GA₃ inactivation. Furthermore bioactive GA₃ deficiency inhibits seed germination by decreasing alpha amylase activity via down regulation of alpha amylase gene expression (Li Liu *et al.*, 2018).

H. Effect of GA₃ on α-amylase Activity

Gibberillic acid is well known to induce the synthesis of α-amylase hydrolysis of starch in rice seeds (Palmiano and Juliano, (1972). Hence the α-amylase activity is more in GA₃ treated seeds. During seed germination, bioactive gas are synthesized in the embryo and transported to the aleurone layer to induce alpha amylase gene expression and alpha amylase synthesis. Then alpha amylase is secreted into the endosperm to hydrolyse the stored starch (Kaneko *et al.*, 2002). Alpha amylase is the major enzyme involved in the hydrolysis of starch to glucose, and accounts of 40-60% of de novo protein synthesis in grains (Sun and Henson, 1991).

I. β-Amylase Activity

A study was done on effect of salinity and GA₃ on the β amylase activity, decrease in the β amylase activity was found under saline treatment but increase in the β amylase under GA₃ treatment and also under combination of NaCl and GA₃ there was an increase in the β amylase activity.

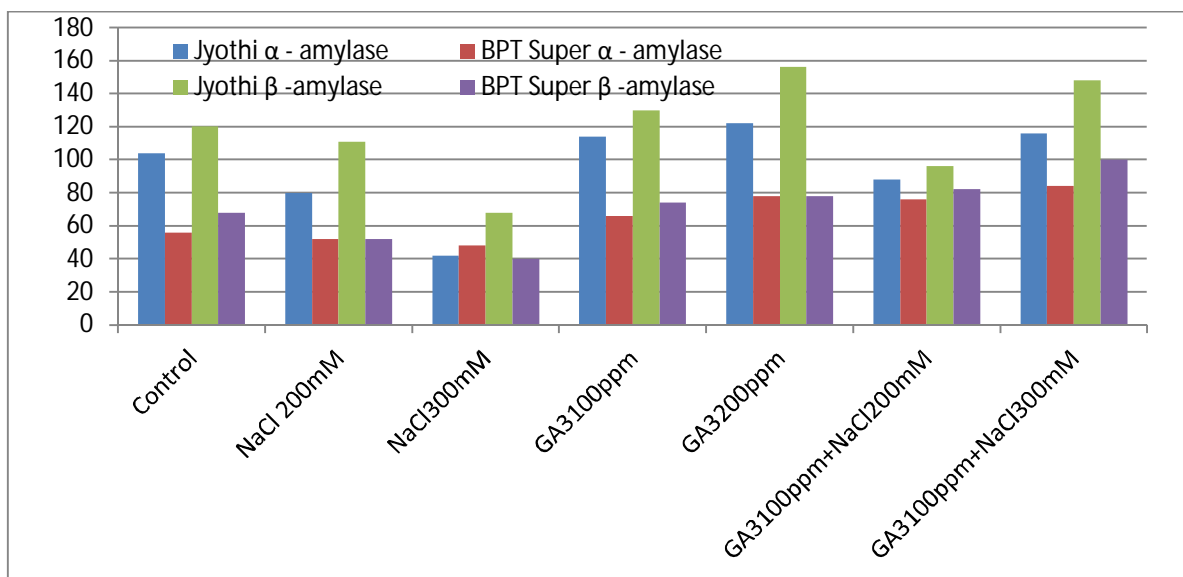
J. Effect of Salinity ON β-amylase Activity

Salinity stress led to a decrease in Beta amylase activity and there was by break down of starch into reducing and non-reducing sugars. Hence the amylase activity was reduced under salinity (Saikat Paul, *et al.*, 2016). Salt stress inhibited the express of β amylase isoenzyme activity (Liu Hua-long *et al.*, 2014).

Table 2: Effect of different concentrations of NaCl, GA₃ and combination of NaCl and GA₃ on α and β amylase (μm/gm) and catalase activity (unit min⁻¹ mg⁻¹protein) of Jyothi and BPT Super paddy cultivars on 14th day.

Cultivar	Parameters	Control	NaCl 200mM	NaCl 300mM	GA ₃ 100ppm	GA ₃ 200ppm	GA ₃ 100ppm + NaCl 200mM	GA ₃ 200ppm + NaCl 300mM
Jyothi	α - amylase	104 ^c	80 ^e	42 ^f	114 ^b	122 ^a	88 ^d	116 ^{ab}
	β -amylase	120 ^c	111 ^c	68 ^e	130 ^b	156 ^a	96 ^d	148 ^a
BPT Super	α - amylase	56 ^d	52 ^{de}	48 ^e	66 ^c	78 ^b	76 ^b	84 ^a
	β -amylase	68 ^d	52 ^e	40 ^f	74 ^{cd}	78 ^{bc}	82 ^b	100 ^a

Means followed by the same letter within a row are not significantly different as indicated by Scheffe (P ≤ 0.05). Significant at P ≤ 0.001.



Graph 2: Effect of different concentrations of NaCl, GA₃ and combination of NaCl and GA₃ on α - amylase and β -amylase content (μm/gm) of Jyothi and BPT Super paddy cultivars on 14th day.

K. Catalase Activity

A test was conducted to study the effect of salinity and GA₃ on the catalase activity. Increase in the catalase activity under the salinity under GA₃ treatment and in combination of NaCl and GA₃ was observed.

L. Effect of Salinity

The amount of H₂O₂ content in leaves gets increased with increasing salinity level than the increase in antioxidative enzyme catalase. Besides CAT detoxifying enzyme is reported to increase under various environmental stresses including salinity (Gueta-Dahan., *et al.*, 1997, Vaidyanathan *et al.*, 2003) and salinity stress increased the CAT activity, was stated by Paul and Roychoudhury. Catalase activity was increased in response to NaCl in rice cultivars with increasing salinity and the results were in concurrence with levels by Kanlaya *et al.*, (2012).

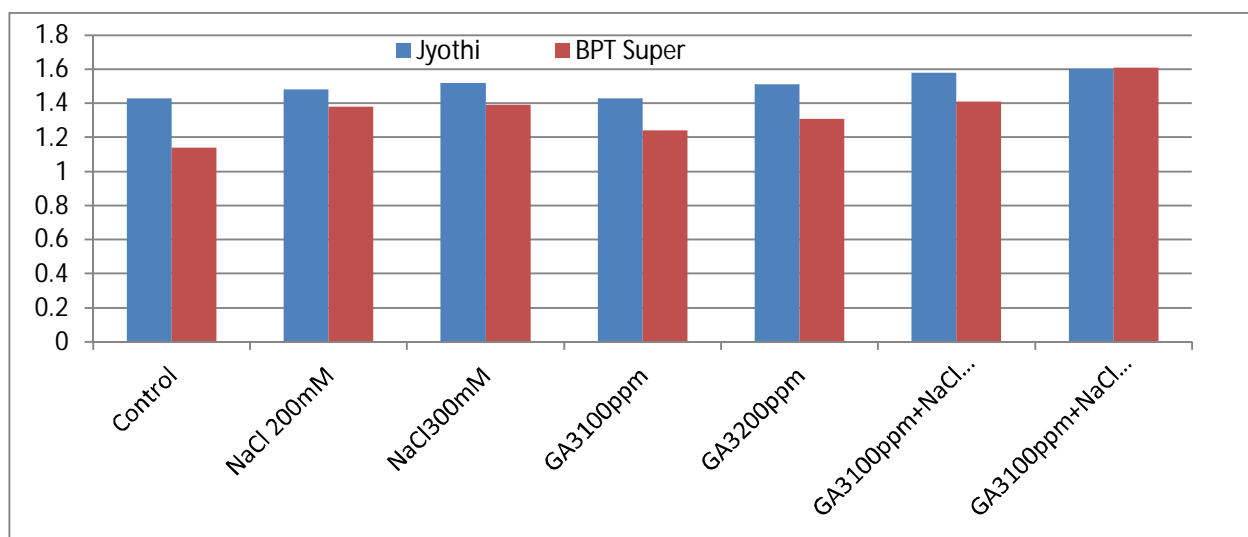
M. Effect of GA₃

The external use of GA₃ increases antioxidant level and will reduce the level of hydrogen peroxide hence the enzyme activity was increased under GA₃ treatment as was reported by Mohammed., *et al.*, (2007)

Table 14: Effect of different concentrations of NaCl, GA₃ and combination of NaCl and GA₃ on Catalase activity (units' min⁻¹ mg⁻¹ protein) of Jyothi and BPT Super paddy cultivars on 14th day.

Cultivar	Control	NaCl 200mM	NaCl 300mM	GA ₃ 100ppm	GA ₃ 200ppm	GA ₃ 100ppm + NaCl 200mM	GA ₃ 200ppm + NaCl 300mM
Jyothi	1.43 ^a	1.48 ^a	1.52 ^a	1.43 ^a	1.51 ^a	1.58 ^a	1.60 ^a
BPT Super	1.14 ^c	1.38 ^b	1.39 ^b	1.24 ^{bc}	1.31 ^{bc}	1.41 ^{ab}	1.61 ^a

Means followed by the same letter within a row are not significantly different as indicated by Scheffe (P ≤ 0.05). Significant at P ≤ 0.001.



Graph 2: Effect of different concentrations of NaCl, GA₃ and combination of NaCl and GA₃ on Catalase activity (units' min⁻¹ mg⁻¹ protein) of Jyothi and BPT Super paddy cultivars on 14th day.

IV. CONCLUSION

α-amylase activity and β-amylase activity was found to be decreased under salinity and the increase was observed under GA₃ condition. Combination of GA₃ and NaCl also increased the α-amylase activity and β-amylase activity, the least affected from salinity was BPT Super cultivar and Jyothi cultivar was more affected. The salinity alleviating role of GA₃ was observed. Catalase activity was increased under salinity in both the cultivars, and salinity was increased under GA₃ treatment and also under the combination of GA₃ and NaCl, so catalase activity was least affected due to the salinity in both the paddy cultivars. With respect to all the above aspects Jyothi paddy cultivar is more salt sensitive compared to the BPT Super, and BPT Super shows better salinity tolerance when compared to the Jyothi cultivar, and salinity alleviating role of GA₃ was observed.

V. ACKNOWLEDGEMENT

The author is grateful to Principal Yuvaraja's College, Constituent autonomous college, University of Mysore, Mysore, Karnataka. For provide necessary facilities used for this work at the department of Botany and Biotechnology. Author also thank to Regional rice research station. V.C. Farm, Mandya.

REFERENCE

- [1] Basu S., gangopadhyay G. and Mukherjee B. 2002. Salt tolerance in rice in vitro: implication of accumulation of Na⁺, K⁺ and proline. *Plant Cell Tiss. Org. Cult.* 69:55-64.
- [2] Beck, E., Ziegler P., 1989. Biosynthesis and degradation of starch in higher plants Briggs, W R (Ed) Annual Review Of Plant Physiology And Plant Molecular Biology, Vol 40 95-118.
- [3] Chakrabarti N, Mukherji S 2003. Alleviation of NaCl stress by pretreatment with phytohormones in *Vigna radiata*. *Biol. Plantarum* 46: 589-594.
- [4] Danai-Tambhale, S., V. Kumar and V. Shriram. 2011. Differential response of two scented indica rice. (*Oryza sativa*) cultivars under salt stress. *J. Stress Physio. & Biochem.* 7(4):387-397.
- [5] Gueta-Dahan Y, Yaniv Z., Silinkas B.A. Ben-Hayyim G. 1997. Slat and oxidative stress: similar and specific responses and their relation to salt tolerance in citrus. *Planta.* 203: 460-499.
- [6] Harinasut P., S. Srinunaka. S. Pitukchaisopola and R. Charoensatapom. 2000. Mechanisms of adaptation to increasing salinity of mulberry: Proline content and ascorbate peroxidase activity in leaves of multiple shoots. *Sci. Asaia*, 2006.
- [7] Hasegawa, P.M., R.A. Bressnan, J.K. Zhu and H.J Bohnert. 2000. Plant cellular and molecular responses to high salinity. *Annual Review of Plant Physiology and Plant Molecular Biology* 51: 463-499.
- [8] Hilal M., Zenoff A.M., Ponessa G., Moreno H. And Massa E.M. 1998: Saline stress alters the temporal patterns of xylem differentiation and alternatiave oxidative expression in developing soybean roots. *Plant Physiol.* 11: 695-701.
- [9] Islam M.S., Hur J.H. and Wang M.H. 2008. The influence of abiotic stresses on expression of zinc finger gene in rice. *Russ J. Pl. P.* 56:695-701.
- [10] ISTA.2009. International seed testing association, News Bulletin. No. 137.
- [11] Jamil M., Iqbal W., Bangash A., Rehman S.U., Imran Q.M. and Rha E.S. 2010. Constitutive expression of OSC3H33, OSC3H50 and OSC3H37 genes in rice under salt stress. *Pak J. Bot.*, 42: 4003-4009.
- [12] Kaneko, M., Itoh, H., Ueguchi-Tanaka, M., Ashikari, M., and Matsuoka, M. 2002. The α -amylase induction in endosperm during rice seed germination is caused by gibberellin synthesized in epithelium. *Plant Physiol.* 128, 1264–1270.
- [13] Kanlaya Kong-ngern, Sumontip Bunnag amd Piyada Theerakalpisut, 2012. Proline, Hydrogen Peroxide, membrane stability and antioxidant enzyme activity as potential indicators for salt tolerance in rice (*Oryza sativa* L.,
- [14] Karrer, E. E., Chandler, J. M., Foolad, M. R., and Rodriguez, R. L. 1993. Correlation between α -amylase gene expression and seedling vigor in rice. *Euphytica*66, 163–169.
- [15] Kim .S. K., T. K. Son, S. Y. Park, I. J. Lee, B. H. Lee, H. Y. Kim and S. C. Lee. 2006. Influences of gibberellin and auxin on endogenous plant hormone and starch mobilization during rice seed germination under salt stress. *Journal of Environmental Biology.* 27(2) 181-186.
- [16] Kumar, V., V. Shriram, Kavi, P. B. Kishor, N. Jawali and M.G. Shitole. 2010. Enhanced proline accumulation and salt stress tolerance of transgenic indica rice by over expressing P5CSF129A gene. *Plant Biotech. Rep.*, 4(1):37-48.
- [17] Linghe Zeng and Michael C. Shannon. 2000. Salinity effects on seedling growth and yield components of Rice. *Crop sci.* 40: 996-1003.
- [18] Liu Hua-long., ShaHan-jing., WangJing-guo., LiuYang Zou De-tang., ZhaoHong-wei, 2014. Effect of Seed Soaking with Exogenous Proline on Seed Germination of Rice under Salt Stress. *Journal of Northeast Agricultural University* 21, (3) 1-6.
- [19] Liu L, Xia W, Li H, Zeng H, Wei B, Han S, Yin C.2018. Salinity Inhibits Rice Seed Germination by Reducing α -Amylase Activity via Decreased Bioactive Gibberellin Content. *Front Plant Sci.* 5(9):275.
- [20] Mansour M.M. and Salama K.H. 2004. Cellular basis of salinity tolerance in plants. *Environ exp. Bot* 52: 113-122.
- [21] Masroor, M., Gautham, C., and Khan, N. 2006. Effect of Gibberllic Acid spary on performance of tomato. *Turk.J. Biol.*, 30:11-16.
- [22] Misratia,K.M. Mohd Razi Ismail, Md Abdul Hakim, Mohamed Hanafi Musa and Adam Puteh. 2013. Effect of salinity and alleviating role of gibberillic acid (GA₃) for improving the morphological, physiological and yield traits of rice varieties. *AJCS* 7(11):1682-1692.
- [23] Mohammed, A. 2007. Physiological aspects of Mungbean plant (*Vigna radiata* L., Wilczek) in response of salt stress and gibberillic acid treatment. *Research Journal of Agriculture and Biological Sciences.*, 3(4):200-2013.
- [24] Munns, R. 2005. Genes and salt tolerance bringing them together. *New phytology*, 167: 645-663.
- [25] Murphy, K.S.T and M.J. Durako. 2003. Physiological effects of short term salinity changes on *Ruppia maritima*. *Aquatic Botany.* 75: 293-309.
- [26] Osakabe Y., Kajita S. and Osakabe K. 2011. Genetic engineering of woody plants: current and future targets in a stressful environment. *Physiol. Plant.* 142: 105-117.
- [27] Palmiano, E. P. and B. O. Juliano: 1972. Biochemical changes in the rice grain during germination. *Plant Physiol.*, 49, 751-756.
- [28] Plaut A., Edelstein M. Ben Hur . 2013. Overcoming salinity barriers to crop production using traditional methods. *Crit. Rev. Plant Sci.* 32(4): 250-291.
- [29] Rahdari, P. and Seyed Meysam Hoseini. 2015. Evaluation of Germination Percentage and Some Physiologic Factors under Salinity Stress and Gibberillic acid Hormone (GA₃) Treatments in Wheat (*Triticum aestivum* L.). *Int. J. Adv. Res. Biol.Sci.* 2(2): 122–131.
- [30] Rahdari, P. S. Tavakoli, and S.M. Hosseini, 2012. Studying stress effect on germination, Proline, Sugar, Protein, Lipid and Chlorophyll content in Purslane (*Portulaca oleracea* L.) leaves. *Journal of Stress Physiology & Biochemistry* 8(1): 182-193.
- [31] Rahman M S., U.A., Haq M.Z., and Gul, S., 2008. Effects of NaCl salinity on wheat (*Titicum aestivum* L) cultivars *World J. Agric. Sci.*, 4(3): 398-403.
- [32] Rogers M.E, Colmer T.D, Frost K, Henry D, Cornwall D, Hulm E, Deretic J, Hughes SR, Craig AD (2008) Diversity in the genus *Melilotus* for tolerance to salinity and waterlogging. *Plant and Soil* 304, 89–101.

- [33] Rogers M.E, Craig A.D, Munns R, Colmer TD, Nichols PGH, Malcolm CV, Barrett-Lennard EG, Brown AJ, Semple WS, Evans PM, Cowley K, Hughes SJ, Snowball R, Bennett SJ, Sweeney GC, Dear BS, Ewing M (2005) The potential for developing fodder plants for the salt-affected areas of southern and eastern Australia: an overview. *Australian Journal of Experimental Agriculture* 45, 301–329.
- [34] Rout N.P., Tripathi S.B. and Shaw B.P. 1997. Effect of salinity on chlorophyll and proline contents in three aquatic macrophytes. *Biol. Plant.* 40: 453-458.
- [35] Roy choudhury A., Basu S., Sengupta D.N. 2011. Amelioration of salinity stress by exogenously applied spermidine or spermine in three varieties of rice differing in their level of salt tolerance. *J. Plant Physiol.* 168(4): 317-328.
- [36] Roy, A.K.D., K. Alam and J. Gow. 2012. A review of the role of property rights and forest policies in the management of the Sundarbans mangrove forest in Bangladesh. *Forest Polciy Econ.* 15: 46-53.
- [37] Saikat paul and aryadeep roychoudhury., 2016. Seed priming with spermine ameliorates salinity stress in the germinated seedlings of two rice cultivars differing in their level of salt tolerance. *Tropical Plant Research.* 3(3): 616-633.
- [38] Santo, C.V. 2004. Regulation of chlorophyll biosynthesis and degradation by salt stress in sunflower leaves. *Science of Horticulure* 103, 93-99.
- [39] Shannon M.C., Rhodes J.D., Draper J.H., Scardaci S.C., Spyres M.D., 1998. Assessment of salt tolerance in rice cultivars in response to salinity problems in California. *Crop Science* 38: 394-398
- [40] Shereen, A., R Ansari., S. Raza., S. Mamta., M.A. Khan and M. Alikhan. 2011. Salinity induced metabolic change in rice (*Oryza sativa* L.,) seeds during germination. *Pak. J. Bot.,* 43(3): 1659-1661.
- [41] Siringam, K., N. Juntawong, S. Chaum and C. Kirdmance 2011. Salt stress induced ion accumulation, ionhomeostasis, membrane injury and sugar contents in salt sensitive rice (*Oryza sativa* L. Spp. Indica) roots under isosmotic conditions. *Afr. J. Biotech.* 10: 1340-1346.
- [42] Sun, Z., and Henson, C. A. 1991. A quantitative assessment of the importance of barley seed α -amylase, β -amylase, debranching enzyme, and α -glucosidase in starch degradation. *Arch. Biochem. Biophys.* 284, 298–305.
- [43] Tavakkoli., E., F. Fatchi, S. Coventry, P. Rengasamy and G.K. McDonald. 2011. Additive effects of Na⁺ and Cl⁻ ions barley growth under salinity stress. *J. Exp.Bot.* 62(6):2189-2203.
- [44] Vaidyanathan H, Sivakumar P, Chakrabarty R, Thomas G. 2003. Scavenging of reactive oxygen speices in NaCl stressed rice (*Oryza sativa* L.,) differential response in salt tolerant and sentive cultivars. *Plant Sci.* 165:1411-1418.
- [45] Yamamoto, A., H. Sawada, I.S. Shim, K. Usui, S. Fujihara, 2011. Effect of salt stress on physiological response and leaf polyamine content in NERICA rice seedlings. *PLANT SOIL ENVIRON.,* 57, 2011 (12): 571–576.
- [46] Yeo A.R. and T.J. Flowers. 1984. Mechanisms of salinity resistance in rice and their role as physiological criteria in plant breeding. P. 151-170. In: *Salinity tolerance in plants strategies for crop improvement* R.C. Staples and G.A. Toennisessen (eds). John Wiley and Sons. New York. USA.
- [47] Zeng, L. And Shannon, M.C. 2000b. Salinity effects on seedling growth and yield components of rice. *Crop Sci.,* 40:996-1003.
- [48] Lowry, O.H., Rosebrough, N.J., Farr, A.L., and Randall, R.J. (1951) *J.Biol.Chem* 193: 265.
- [49] Moore, S. and Stein, W. H., *J. Biol. Chem.,* 176, 367 (1948).



10.22214/IJRASET



45.98



IMPACT FACTOR:
7.129



IMPACT FACTOR:
7.429



INTERNATIONAL JOURNAL FOR RESEARCH

IN APPLIED SCIENCE & ENGINEERING TECHNOLOGY

Call : 08813907089  (24*7 Support on Whatsapp)